

Biocultural Implications of Oral Pathology in an Ancient Central California Population

Mark C. Griffin*

Department of Anthropology, San Francisco State University, 1600 Holloway Avenue, San Francisco, CA 94132

KEY WORDS bioarchaeology; dental caries; periapical abscess; dental wear

ABSTRACT Bioarchaeologists have long noted two unusual trends in the dentitions of prehistoric Native Californian populations: high rates of wear and low prevalence of caries. The Central California site of CA-CCO-548 offers a unique opportunity to examine the relationship between oral pathology and extreme dental wear in a large ($n = 480$), ancient (4,300–3,100 BP), and temporally well-defined population sample. This study specifically examines three interrelated processes of the oral cavity in this population: dental wear, dental caries, and periodontal disease. The results show high levels of dental wear (average of 6.1, Smith system), low frequencies of carious lesions (2.5%), low frequencies of periodontal disease (17.8%), and high frequencies of periapical abscesses (10.7%). The pathological processes examined here have complicated mul-

tifactorial etiologies. However, they all share the common primary etiological agents of facultative pathogenic bacteria proliferation in the oral biofilm. Integration of the current etiological explanations for infections of the oral cavity, information from the ethnographic record pertaining to subsistence and activity patterns in Native Californian populations, and statistical analysis of specific disease and wear patterns leads to a novel explanation for the observed pattern of oral pathology in this population sample. Specifically, the introduction of antibacterial compounds through dietary items and non-alimentary tooth use is suggested as the most likely explanation for the unusually low prevalence of dental caries and periodontal disease. *Am J Phys Anthropol* 154:171–188, 2014. © 2014 Wiley Periodicals, Inc.

Two pervasive features noted among precontact California Native American populations are extreme dental wear (Leigh, 1928; Molnar, 1971a and b; Walker, 1978; Reinhardt, 1983; Jurmain, 1990; Bartelink, 2006; Grant, 2010; Blake, 2011) and remarkably low prevalence of dental caries (Schulz, 1981; Walker and Erlandson, 1986; Hoffman, 1987; Jurmain, 1990; Bartelink, 2006). Precontact individuals from California sites routinely exceed the highest score for the various dental wear systems (Reinhardt, 1983; Jurmain, 1990). Reinhardt (1983) noted that dental wear in precontact California individuals is so extreme that it often obliterates the crown of the tooth. This creates a condition where the tooth roots are functioning in occlusion. Individuals exhibiting this wear pattern continued to masticate using the remaining root stubs, frequently wearing them down to the alveolar margin. Interpretations vary as to the causative factors involved in this extreme wear. Explanations include the unintentional inclusion of grit and stone from food processing (Jurmain, 1990), food sources with natural abrasive inclusions (Walker, 1978), and non-alimentary dental wear from producing baskets and cordage (Molnar, 1971b, 1972; Schulz, 1977; Grant, 2010; Blake, 2011).

Extraordinarily low prevalence of dental caries is also a ubiquitous feature of precontact California Native populations (Schulz, 1981; Walker and Erlandson, 1986; Jurmain, 1990; Bartelink, 2006). This observation is remarkable considering the shift in many parts of California to a carbohydrate-rich, plant-intensive economy in the Middle Holocene (Rosenthal et al., 2007). Precontact California populations never adopted agriculture as was seen in many parts of precontact North America. The transition in California was more of a move from hunting-gathering to a pattern of foraging and “tending.”

That is, while agricultural plants were never adopted in California, the careful management of acorn producing oak groves was a dominant feature of the culture beginning around 4500 BP (Basgall, 1987; Beaton, 1991; McCarthy, 1993; Rosenthal et al., 2007). The catalyst for this transition was likely the increasing aridity of the environment caused by the Altithermal. The increasing aridity would have in turn caused populations that retained a hunter-gatherer economy to exceed the carrying capacity of their environment (Basgall, 1987). Acorns provided a more reliable alternative to other available resources (Tushingham and Bettinger, 2013).

Extreme wear has been suggested as a possible contributory cause for the low prevalence of caries in precontact California populations (Jurmain, 1990; Bartelink, 2006), as well as low caries prevalence observed in populations outside of California (Powell, 1985; Maat and Van der Velde, 1987; Kerr et al., 1990; Newman, 1999; Caglar et al., 2007; Buzon and Bombak, 2010). Researchers however, have come to differing conclusions regarding the relationship between wear and caries. Some researchers have suggested that the elimination of occlusal fissures on the posterior dentition

Abbreviations: AC, Alveolar crest; CEJ, Cemento-enamel junction; LMW, Low molecular weight

*Correspondence to: Dr. Mark C. Griffin, Department of Anthropology, San Francisco State University, 1600 Holloway Avenue, San Francisco, CA 94132. E-mail: mgriffin@sfsu.edu

Received 10 October 2013; accepted 29 January 2014

DOI: 10.1002/ajpa.22491

Published online 12 February 2014 in Wiley Online Library (wileyonlinelibrary.com).

through excessive dental wear reduces the prevalence of caries (Powell, 1985; Newman, 1999). These fissures are a primary location of the oral biofilm, which in turn is one of the essential etiological components of caries (Macek et al., 2003; Ferreira Zandona et al., 2012). Other research has indicated a synergistic relationship between wear and caries (Hardwick, 1960; Miles, 1969; Silverstone et al., 1981). Increased attrition has the capacity to expose the softer dentin to cariogenic bacteria and therefore predispose individuals to higher frequencies of carious lesions. Alternatively, Meiklejohn and coworkers (1992) concluded that there is no causal relationship between caries and dental wear. They suggest that the processes are independent of one another.

The aim of this study is to examine the relationship between extreme dental wear and oral pathology in the ancient California population sample from CA-CCO-548. This will be done by assessing the prevalence of dental caries and periodontal disease in this sample and statistically comparing prevalence of these oral pathologies to levels of dental wear. Patterns of oral pathology are assessed in the context of recent advances in the understanding of oral disease etiologies. CA-CCO-548 offers the unique opportunity to examine the potential interaction between extreme dental wear and caries prevalence in a well-documented precontact California population. Research at the site has yielded extensive information pertaining to skeletal biology, paleobotany, zooarchaeology, material culture, and environmental context (Grant, 2010; Wiberg, 2010; Guidara, 2012; Eerkens and Bartelink, 2013). Based on previously reported patterns of dental wear and caries in precontact California, it is hypothesized that there will be a statistically significant relationship between high levels of wear and low prevalence of dental caries.

ORAL PATHOLOGY AND DENTAL WEAR

It is estimated that only 10% of the cells in the human body are mammalian (Marsh et al., 2011). The remaining 90% are primarily comprised of the resident human microflora that live in and on our body (Wilson, 2005). Most of the nearly 1200 indigenous species of microflora that inhabit the human oral cavity are commensal (Jenkinson, 2011). That is, most of our oral microfloras are not harmful and some are actually beneficial. These microorganisms live in normally stable microbial communities of viruses, mycoplasma, bacteria, Archaea, fungi and protozoa collectively referred to as biofilm (Meurman et al., 2004; Marsh and Martin, 2009; Palmer, 2014). A biofilm is a three-dimensional structured microorganism community with a complex web of fluid channels for transport of substrate, waste products, and signal molecules (Costerton, 1999; Scheie and Peterson, 2004). The oral microflora are initially introduced through salivary transmission associated with intimate human contact and establish themselves on the hard, non-desquamating surfaces of the oral cavity by 19–31 months of age (Caufield et al., 1993). In the normal individual there is a delicate balance between commensal and facultative pathogenic microflora of the oral cavity (Liljemark and Bloomquist, 1996; Kidd and Fejerskov, 2004; Palmer, 2014). Oral microorganisms dynamically adapt to changes in their immediate environment by using various mechanisms such as altering their gene expression (Fornig et al., 2000). This capacity allows these protean communities to maintain a synergistic

homeostasis. Disruptions of the oral environment that exceed this adaptive capability allow facultative pathogenic species to proliferate resulting in disease processes. Once the disease process commences, pathogenic microorganisms have the capacity to make the process self-sustaining. Pathogenic oral microorganisms increase their fitness advantage over commensal microflora by using mechanisms like altering their gene expression, regulating conditions of the local environment, and releasing metabolic products that kill or suppress competitive bacterial species (Merritt and Qi, 2012; Corcuera et al., 2013; Koo et al., 2013). Two disease processes that are the result of this type of microbial disruption are dental caries and periodontal disease.

Caries etiology

Dental caries results from three interrelated contributors: (1) disturbance of the balance between commensal and facultative pathogenic oral microflora, (2) frequent consumption of fermentable carbohydrates, and (3) host susceptibility mediated by genetic and non-genetic influences (Lingström et al., 1994; Marsh, 1995). There are more than forty different species of *Streptococcus* (Innings et al., 2005), many of which colonize the human oral cavity shortly after birth (Law et al., 2007). Many of these species are commensal, some are pathogenic. Seven pathogenic species of *Streptococcus* have been implicated in human caries and endocarditis (Coykendall, 1989). Collectively these are known as the mutans streptococci. The two most common mutans streptococci in humans are *Streptococcus mutans* and *Streptococcus sobrinus* (Coykendall, 1989). Although mutans streptococci are the most frequently implicated in caries etiology, research demonstrating the frequent presence of caries in the absence of these organisms indicates that it is the disturbance of oral microbiotic balance that is more important in caries etiology than the presence of a particular organism (Kleinberg, 2002; Giacaman et al., 2010; Simón-Soro et al., 2013). When pathogenic bacteria proliferate in the oral cavity and consume carbohydrates, they produce metabolic wastes which contribute organic acids and proteases to the oral fluids (Liljemark and Bloomquist, 1996; Featherstone, 2004). These metabolic products make the oral environment more favorable for the pathogenic organisms by lowering pH and in sufficient concentration, can dissolve the mineral component of dental enamel producing a carious lesion. Left unchecked, proteolytic decay can completely destroy a tooth and enter the circulatory system exposing extra-oral sites to infection. Oral bacteria, especially those connected with dental caries, have been shown to have a major role in the development of serious extra-oral diseases including cardiovascular disease, rheumatoid arthritis, inflammatory bowel disease, colorectal cancer, and respiratory tract infections (Meurman et al., 2004; Han and Wang, 2013).

Periodontal disease etiology

Periodontal diseases are multifactorial infections of the sulcular region between the tooth surface and the gingival margin (Holt and Ebersole, 2005). At least 400 of the more than 1200 bacterial species that colonize the oral cavity inhabit subgingival sites (Paster and Dewhirst, 2009). Up to 150 different species are present in any individual's subgingival plaque (Buduneli and Kinane, 2011). Similar to dental caries etiology, most of the

bacterial species in subgingival biofilms are commensal. Fewer than twenty are considered to be major periodontal pathogens (Socransky and Haffajee, 2005). The initiation of periodontal disease commences with a shift in subgingival bacterial colonies from ones predominated by gram-positive bacteria to communities dominated by anaerobic gram-negative bacteria (Slots, 2010). The triggers that cause the overgrowth of these normally commensal species and the virulence determinants associated with them are not understood (Holt and Ebersole, 2005).

The pathogenic bacterial species primarily implicated in periodontal diseases are *Tannerella forsythia*, *Porphyromonas gingivalis*, *Aggregatibacter actinomycetemcomitans* (Laine et al., 2013). The predominance of these bacterial species has been clearly linked to chronic periodontal disease (Dentino et al., 2013). Tissue destruction resulting from chronic periodontal disease is due to collateral damage from the host immune response rather than destruction directly caused by the pathogenic bacteria (Taubman et al., 2005; Dentino et al., 2013; Cekici et al., 2014). The overgrowth of pathogenic species stimulates the release of host produced inflammatory cytokines, chemokines, and mediators (Silva et al., 2007). Host leukocytes responding to infection do not discriminate between the invading bacteria and host tissue. The result of this failure to discriminate pathogen from host tissue is the collateral destruction of gingival and skeletal tissues adjacent to the infection site (Liu et al., 2010; Preshaw and Taylor, 2011). With continued infection there is progressive destruction of gingiva, the periodontal ligament, and the underlying alveolar bone tissue. Unabated, periodontal disease will cause sufficient alveolar recession to result in tooth exfoliation (Oliver and Brown, 1993; Thomson et al., 2013). The bacteria associated with periodontal disease have also been linked to increased risk for cardiovascular diseases (Kebshull et al., 2010; Reyes et al., 2013; Schenkein and Loos, 2013) and other serious extra-oral infections (Han and Wang, 2013).

Periapical periodontitis etiology

More than 450 bacterial species have been identified in endodontic infections (Siqueira and Rôças, 2009). This is by far the most diverse assemblage of oral pathogens in all of the diseases of the oral cavity. Most endodontic infections are polymicrobial, involving various combinations of the most prevalent bacterial species (Brauner and Conrads, 1995; Dahlén, 2002). These include microaerophilic streptococci (*S. anginosus*, *S. constellatus*, *S. intermedius*), anaerobic streptococci (*Peptostreptococcus anaerobius*, *P. micros*), gram-positive anaerobic rods (*Eubacterium* spp., *Actinomyces* spp., *Propionibacterium* spp.), and gram-negative anaerobic rods (*Porphyromonas* spp., *Prevotella* spp., *Bacteroides* spp., *Campylobacter* spp., *Fusobacterium* spp., *Treponema* spp.). Endodontic infection involves an opportunistic infection of the dental root canal system (Siqueira, 2002). Infection of the root canal requires the absence of vital pulp tissue (Siqueira and Rôças, 2009). Pulp tissue deficiency results from either pulp necrosis which occurs as part of a disease sequela (caries, trauma, periodontal disease) or pulp removal as part of a medical procedure (Abbott, 2004). In response to the proliferation of pathogenic microorganisms, the body mounts an immune response involving the same inflammatory cytokines, chemokines, and

mediators found in other periodontal diseases (Silva et al., 2007). The resulting abscess is a localized collection of the immune responders and disintegrated tissues (pus). Drainage of the cellular debris is accomplished through the formation of a fistula. In the case of periodontal abscesses, the drainage is most likely through the periodontal pocket because that is the path of least resistance (Dahlén, 2002). The sequelae of chronic infection include tooth exfoliation as well as bacterial diffusion to sinuses and other facial spaces to form cellulitis (Siqueira and Rôças, 2009).

Dental wear etiology

The age-related phenomenon of dental enamel reduction (dental wear) occurs in all human populations and is recognized as a contributing factor in oral disease risk (Lee et al., 2012). Dental wear results from three contributors: (1) attrition results from tooth-on-tooth contact, (2) abrasion is produced by the friction of foreign substances on the enamel, and (3) erosion is the chemical dissolution of enamel not due to biofilm (Kaidonis et al., 1998). Dental wear is dependent on two major variables, diet and age of the individual. Dental wear in populations that have a more abrasive diet (e.g., hunter-gatherer subsistence) tends to be more rapid and severe than in those that have a softer, more highly processed diet (e.g., agricultural subsistence) (Deter, 2009). Not only does subsistence system affect the overall level of wear but it also has an effect on the specific pattern of wear on the teeth. Given the fibrous nature of many hunter-gatherer diets, the wear pattern generally tends to be flat across the occlusal surface (Smith, 1984). The softer texture of the agricultural diet tends to produce a "scooped out" appearance to the occlusal surface (Smith, 1984). Severe wear or wear resulting from specific, repetitive alimentary and non-alimentary activities can result in wear patterns that significantly deviate from these generalized patterns (Kieser et al., 2001; Kaifu et al., 2003).

The pathological and non-pathological physiological processes discussed above all have the capacity of altering the oral environment. Recent clinical research indicates that pathological processes in the oral cavity are the result of disturbances in the normal balance of our native microbiotic communities rather than invasion by exogenous organisms. Dental wear also has the capacity of altering the balance of the microbiotic community through both disruption and facilitation. That is, dental wear in general disrupts the microbial environment through biofilm removal, while excessive wear offers pathological species access to normally protected areas. Based on the etiological foundations of the four processes examined here, it is hypothesized that high levels of dental wear will correspond to low prevalence of oral pathology in the CA-CCO-548 population sample. Specifically, the consistent removal of biofilm through heavy dental wear should prevent the proliferation of pathogenic microorganisms in the oral cavity.

MATERIALS AND METHODS

The population sample

The population sample examined in this study derives from CA-CCO-548, an ancient Native American mortuary site in the Central Valley of California. Human occupation of the Central Valley can be traced back to

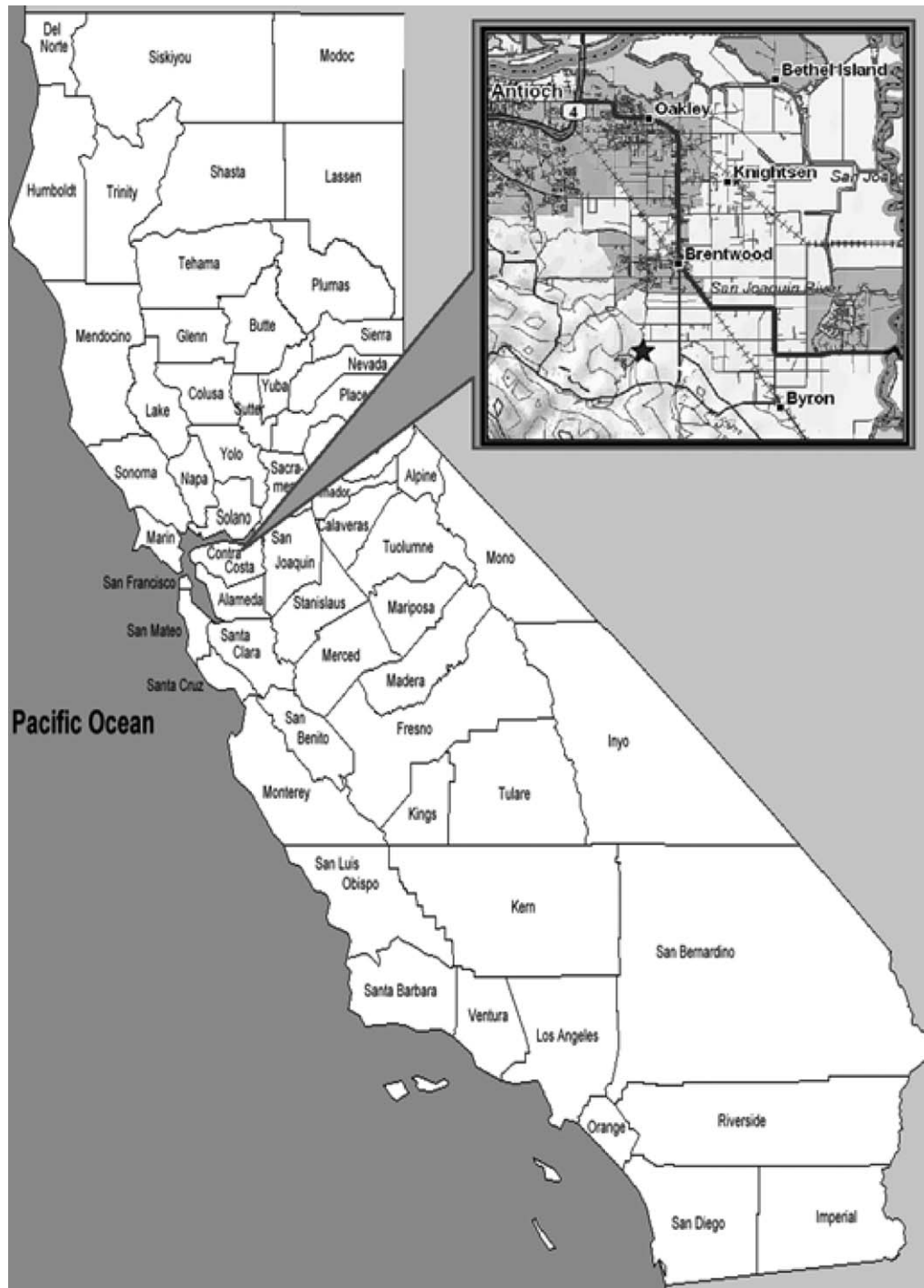


Fig. 1. Geographic location of the Vineyards Site (CA-CCO-548).

around 13,500 BP (West et al., 1991). Occupation during the Paleo-Indian (13,500-10,500 BP) and Lower Archaic (10,500-7,500 BP) is represented by isolated finds (Rosenthal et al., 2007). Occupation in the valley during the Middle Archaic (7,500–2,500 BP) is represented by numerous archaeological sites scattered across the entire region. Despite the presence of numerous sites, extensive human skeletal samples from the Early Period of California (11,000–3,000 BP) are rare (Walker, 2006). This trend holds true for the early Middle Archaic of the

Central Valley (ca. 7,500–4,000 BP) where few cultural deposits have been found due to obliteration or burial by alluvial action (Rosenthal et al., 2007).

The archaeological deposits at CA-CCO-548 represent a rare example of a stratified, multi-component site from the Middle Archaic (Fig. 1). The human skeletal sample, comprised of at least 480 individuals, is the largest for this pivotal time period in California. Geoarchaeological analysis has identified four discrete stratigraphic temporal components for the site, spanning a time from 7000

TABLE 1. Age and sex distribution for CA-CCO-548

	0-5	6-10	11-15	16-20	21-25	26-30	31-35	36-40	41-45	46-50	51-55	56-60	Total
Female	0	0	1	2	6	2	8	15	16	25	5	0	80
Male	0	0	0	1	4	10	14	24	27	28	3	0	111
Indet.	12	11	6	5	8	14	14	14	24	26	9	1	144
Total	12	11	7	8	18	26	36	53	67	79	17	1	335

TABLE 2. Frequencies of antemortem tooth loss (AMTL) and postmortem tooth loss (PMTL) in maxillary and mandibular dentitions

	Total sample			Female			Male		
	N	% AMTL	% PMTL	N	% AMTL	% PMTL	N	% AMTL	% PMTL
Maxilla									
I1	252	9.1	26.2	79	3.8	34.2	116	16.4	21.5
I2	273	4.8	20.1	88	3.4	21.6	128	7.8	19.5
C	293	2.4	9.2	91	0.0	11.0	133	5.3	6.8
P3	289	3.1	5.9	90	2.2	3.3	136	5.1	4.4
P4	291	7.9	5.1	89	7.9	2.2	132	9.8	6.8
M1	296	11.5	0.7	86	5.8	1.1	128	19.5	0.0
M2	294	6.8	2.0	89	3.4	3.4	135	11.1	2.2
M3	239	7.9	8.4	68	4.4	11.8	120	10.0	7.5
Total	2227	6.6	9.4	680	3.8	10.7	1028	10.5	8.4
Mandible									
I1	263	8.4	19.8	82	11.0	20.7	122	8.2	16.4
I2	301	5.3	14.3	96	5.2	16.7	139	6.5	11.5
C	331	3.3	15.7	96	7.3	16.7	151	2.6	15.2
P3	343	4.9	10.5	102	9.8	7.8	152	3.9	12.5
P4	361	6.1	6.9	112	11.6	2.7	163	6.5	7.4
M1	379	6.1	4.2	112	13.4	5.3	164	3.6	0.6
M2	373	7.5	1.9	114	14.0	1.7	168	7.1	1.2
M3	307	11.4	7.5	85	16.5	12.9	149	11.4	4.0
Total	2658	6.5	9.5	799	11.1	9.9	1208	6.2	8.2
Site total	4885	6.6	9.5	1479	7.8	10.3	2236	8.2	8.3

to 3100 cal BP (Meyer, 2010). All but five of the burials were recovered from the components dating between 4,300 and 3,100 cal BP. The majority ($n = 304/480$) of the mortuary features were from the component that dates from 3,600 to 3,400 cal BP. Sex determination could be ascertained for 191 individuals (Table 1). Among the individuals for which sex could be determined, 41.9% ($n = 80/191$) are female and 58.1% ($n = 111/191$) are male (Griffin et al., 2010). Age at death could be estimated for 335 individuals (Table 1). Subadults account for 10.1% ($n = 34/335$) of the sample for which age-at-death could be estimated and 89.9% ($n = 301/335$) are adults (Griffin et al., 2010). It is notable, especially for the pathological conditions that are age-progressive, that 49% ($n = 164/335$) of the sample are older than 40 years of age and 29% ($n = 97/335$) are older than 45 years of age.

Carious lesions

To examine frequencies of carious lesions in the sample from CA-CCO-548, each tooth was examined with 3.5× dental loupes, a 3-mm Microlux Transilluminator™ fiber optic diagnostic light, and stainless steel dental probes. Fiber optic transillumination offers a superior method of lesion detection compared to other macroscopic methods (Davies et al., 2001; Pretty, 2006). Carious lesions were only counted with the presence of a definable pit (cf., Lukacs, 1989; Buikstra and Ubelaker, 1994; Cucina et al., 2011; Halcrow et al., 2013). Tooth discolorations alone were not counted as carious lesions. Wasterlain and coworkers (2011) suggest that this method underestimates the actual number of carious

teeth, however, all of the comparative studies used here use the minimum criteria of a definable pit for lesion identification. Lesions were recorded with regard to their location on the tooth (occlusal, cervical, buccal, lingual, mesial, and distal). Maximum lesion size was measured in millimeters using a Paleo-Tech Helios type needle point caliper. The data reported here have been reduced to lesion presence and absence because the comparative studies for the region do not include details regarding numbers of lesions on particular teeth, lesion sizes, or lesion locations. To facilitate comparison with other sites, left and right antimeres were pooled while maintaining separate counts for each tooth classification in the maxillary and mandibular dentitions. Maintaining counts for separate tooth classifications allows for the detection of intra-oral patterns of disease (cf., Hillson, 2001; Caglar et al., 2007; Temple and Larsen, 2007; Wasterlain et al., 2009; Cucina et al., 2011). Correction factors to account for antemortem tooth loss have been suggested for the examination of caries rates (e.g., Lukacs, 1995; Duyar and Erdal, 2003). Correction factors are not used here because none of the comparative studies used such factors. Antemortem tooth loss (Table 2) is very low in this population sample (overall 6.6%; $n = 322/4885$) and there are no significant differences in AMTL between sexes.

Periodontal disease and periapical abscesses

Clinical researchers that have measured alveolar bone loss in dry skulls consider distances of 1–3 mm from the cemento-enamel junction (CEJ) to the alveolar crest



Fig. 2. Examples of teeth with stage 9 wear.

(AC) to be normal (Davies et al., 1969; Tal, 1985). Most bioarchaeological studies consider CEJ to AC measurements in excess of 2 mm to be potentially pathological (Clarke et al., 1986; Wasterlain et al., 2011; DeWitte, 2012). Bioarchaeological research has shown that accurate documentation of periodontal disease in skeletal populations depends on CEJ to AC measurement and corroborating inspection of the AC for pathologic activity (Klaus and Tam, 2010; Wasterlain et al., 2011; Vodanović et al., 2012). This method accounts for the phenomenon of continuous eruption. The human dentition continues to erupt beyond the time when teeth initially come into occlusion (Tal, 1985; Whittaker et al., 1985; Kaifu et al., 2003; Haytac et al., 2013). The process is age-progressive and is not directly dependent on other factors such as wear (Varrela et al., 1995), although excessive wear can accelerate the process (Newman, 1999). For this study, CEJ to AC measurements were taken for each tooth and the adjacent AC was macroscopically examined for porosity. Teeth with CEJ to AC measurements in excess of 2 mm that also exhibited adjoining alveolar bone porosity were considered pathological.

Periapical abscesses can be associated with periodontitis (periodontal abscess); however, the vast majority of periapical abscesses are of endodontic origin (Dahlén, 2002). Other research has suggested that most periapical cavities in the archaeological record may be the result of periapical granuloma and cysts (Dias and Tayles, 1997). This view is not supported by the clinical research that indicates that granuloma and cysts are relatively rare, accounting for only 15% of periapical lesions (Shear, 1992; Nair, 1997; Nair, 2003). Differential diagnosis of the two conditions is relatively straightforward. Granuloma and cysts are closed pathological cavities (Nair, 2003) that measure less than 3 mm (Dias and Tayles, 1997). Periapical abscesses are a localized collection of pus with a well-defined sinus tract (fistula) which generally exits through the periodontal pocket (Dahlén, 2002) and measure greater than 3 mm (Dias and Tayles, 1997).

To assess the frequency of periapical abscesses, each tooth position was examined macroscopically for the presence of periapical alveolar defects. Defects were recorded by specific location and size. Defects that measured in excess of 3 mm and exhibited a well-defined sinus tract were considered to be periapical abscesses.

For purposes of intersite comparison, left and right maxillary and mandibular tooth positions were pooled.

Dental wear

While the systematic observation of dental wear has long been a standard part of bioarchaeological studies, the method of measurement has been far from standardized. Various scoring systems have been proposed attempting to account for variations in dentin exposure (Molnar, 1971b; Scott, 1979; Molnar et al., 1983; Richards, 1984; Dreier, 1994), reduction of crown height (Tomenchuk and Mayhall, 1979), and angle of wear plane (Molnar, 1971b; Hall, 1976; Smith, 1984). Unfortunately, standardizing a method of recording dental wear is nearly impossible because the way in which human dentitions wear is far from standard and depends on a number of complex biological and behavioral factors.

Each tooth in the CA-CCO-548 sample was scored for dental wear using the Smith wear system (Smith 1984). The Smith system was used here because of its simplicity, which reduces inter- and intra-observer error. A considerable number of teeth in precontact California Native populations exceed stage eight in the system. Following the recommendation of Reinhardt (1983), a stage nine was added for this study. Stage nine was used when the entire crown of the tooth was worn away as well as a considerable portion of the root (Fig. 2). To facilitate comparison with data from other studies, left, and right antimeres were pooled while maintaining separate counts for each tooth classification in the maxillary and mandibular dentitions. Pooling the antimeres is less than ideal for accurately recording specific wear patterns. However, published comparison data for populations of appropriate geographic location and time period mostly consists of pooled data.

RESULTS

Examination of carious lesion prevalence in the CA-CCO-548 sample (Tables 3 and 4) reveals that the overall lesion frequency is quite low. The frequency of teeth with carious lesions is 2.5% for the maxillary dentition ($n = 44/1782$) and 2.6% for the mandibular dentition ($n = 56/2175$). Compared with other population samples from Central California (Tables 3 and 4) the overall frequencies of carious lesions in this population sample are

TABLE 3. Carious lesion frequencies in Central California sites

Tooth	CCO-548		SJO-17 ^a		SJO-154 ^b		SRI-41 ^c		SRI-2A ^c		SRI-2B ^c	
	N	%	N	%	N	%	N	%	N	%	N	%
Maxilla												
I1	153	0.6	36	2.8	26	7.7	82	1.2	16	0	36	0
I2	190	0	35	2.8	32	3.1	107	6.5	20	0	54	0
C	236	0.8	52	7.7	35	14.3	124	16.9	22	0	77	2.6
P3	248	0.4	59	7.7	32	9.4	111	16.2	16	0	73	0
P4	237	0.4	71	1.4	32	12.5	108	6.5	19	5.3	70	11.4
M1	253	6.7	59	23.7	29	17.2	103	28.2	17	35.3	93	8.6
M2	263	2.3	62	14.5	24	8.3	101	29.7	19	31.6	85	17.6
M3	202	7.9	43	14	21	33.3	70	34.3	14	28.6	43	18.6
Total	1782	2.5	417	9.3	231	12.5	806	17	143	11.9	531	7.7
Mandible												
I1	186	0.5	33	0	30	3.3	105	0	8	0	39	0
I2	231	0.4	34	0	33	6.1	126	1.6	12	0	46	0
C	258	0	38	7.9	33	6.1	129	3.9	18	5.6	52	0
P3	276	0.7	39	2.6	39	15.4	126	7.9	15	6.7	53	0
P4	296	0	40	0	40	10	120	10.8	16	0	52	0
M1	340	1.8	45	4.4	35	14.3	107	22.4	14	14.3	63	12.7
M2	339	5.3	41	4.9	32	15.6	118	19.5	14	21.4	53	7.5
M3	249	11.2	41	17.1	33	30.3	81	17.3	11	27.3	45	13.3
Total	2175	2.6	311	4.8	275	12.7	912	10	108	9.3	403	4.5
Site Total	3957	2.5	728	7.4	506	12.6	1718	13.3	251	10.8	934	6.3

^a Molnar 1971b.
^b Hoffman 1987.
^c Walker and Erlandson 1986.

extraordinarily low. Furthermore, the combined lesion frequencies for CA-CCO-548 are more than 50% lower than any of the other comparative population lesion frequencies. Clinical research has established that there is a general gradient of caries risk which progressively increases from anterior tooth positions to posterior ones (Afroughi et al., 2010; Sheiham and Sabbah, 2010). Second molars consistently have the highest frequencies of carious lesions due to their complex fissure patterns (Ferreira Zandona et al., 2012). Given these relationships, it is important to establish that combined lesion frequencies are not the result of over-representation of one tooth type when reporting overall frequencies of carious lesions. The difference in proportion of anterior teeth to posterior teeth in this sample is statistically significant ($\chi^2 = 111.37$, P -value = 0.0001). However, considering that the higher proportion of teeth is contributed by the posterior dentition (68.3%; $n = 2703/3957$) and the highest proportion of a tooth class is contributed by just the molars (41.6%; $n = 1646/3957$), the low overall lesion frequencies become more notable. With a high proportion of molar teeth, one would expect resulting higher overall frequencies of lesions.

Considering the molar teeth separately, the overall low frequencies of carious lesions in this sample become more remarkable. The frequency of carious lesions in maxillary molars is 5.4% ($n = 39/718$) and 5.6% ($n = 52/928$) for mandibular molars. These are far lower than the frequencies reported for the molar teeth in most of the comparative samples. The only comparative site with frequencies this low is the Channel Islands site of CA-SRI-2B, reporting frequencies of 9.5% ($n = 21/221$) for the maxillary molars and 5.6% ($n = 28/501$) for the mandibular molars. The frequencies of molar lesions in the CA-CCO-548 sample are less than half that observed for most other sites and less than one quarter of that seen at some of the comparative sites.

When assessing frequencies of carious lesions, the age-progressive nature of the disease is always a concern. This is especially true of the molars. The age structure of CA-CCO-548 is of particular concern given that 49% of the sample are estimated to be older than 40. Table 5 shows the distribution of carious lesions in seven age groups for the molar series. The differences in frequency of carious lesions between the age groups does not show a consistent age progression and are not statistically significant ($\chi^2 = 9.699$, P -value = 0.138). Similar to previous research in other population samples, females in the CA-CCO-548 sample exhibit higher frequencies of carious lesions for most teeth (Fig. 3). However, only two teeth (lower M2 and M3) have differences between males and females that are statistically significant ($\chi^2 = 7.602$, P -value = 0.022; $\chi^2 = 7.897$, P -value = 0.006).

Examination of the average wear across the entire CA-CCO-548 sample (Fig. 4) indicates a population exhibiting considerable dental wear. The average overall dental wear score for the sample is 6.1 with more than 30% of the sample exhibiting average wear scores of eight or above. Comparing composite wear scores for samples can be somewhat deceiving when individuals are wearing their teeth in unusual ways. The mean wear separated by tooth class shows that the overall wear in this population sample is relatively uniform in terms of degree of wear (Fig. 5). Comparison of male and female average wear scores (Fig. 5) reveals very little difference in wear levels between the two sexes. None of the differences are statistically significant. A comparison of CA-CCO-548 with two other Central California pre-contact sites for average wear scores in each tooth class of the upper and lower dentition (Fig. 6) reveals a striking similarity in amount and pattern of wear. All three sites exhibit considerable wear, with the highest wear values on the upper and lower first molars.

Comparing frequencies of the three separate pathological conditions of the periodontal region reveals a

TABLE 4. Carious lesion frequencies in females and males for Central California sites

Tooth	CCO-548						SJO-17 ^a						SJO-154 ^b						SRI-41 ^c						SRI-2A ^c						SRI-2B ^c					
	Female		Male		%		Female		Male		%		Female		Male		%		Female		Male		%		Female		Male		%		Female		Male		%	
	N	%	N	%	N	%	N	%	N	%	N	%	N	%	N	%	N	%	N	%	N	%	N	%	N	%	N	%	N	%	N	%				
Maxilla	40	2.5	69	0.0	14	0.0	22	4.5	11	18.2	9	0	42	2.4	40	0.0	11	0.0	5	0.0	22	0.0	22	0.0	14	0.0	22	0.0	31	0.0	23	0.0				
I1	60	0.0	87	0.0	13	7.7	22	0.0	14	7.1	12	0	51	9.8	56	3.6	12	0.0	8	0.0	31	0.0	31	0.0	44	0.0	44	0.0	33	6.1	36	0.0				
I2	73	0.0	108	1.9	23	13.0	29	3.4	17	23.5	15	6.7	59	20.3	65	13.8	14	0.0	8	0.0	44	0.0	44	0.0	37	0.0	37	0.0	36	6.3	32	6.3				
P3	76	1.3	112	0.0	20	10.0	32	9.4	17	11.8	12	8.3	55	20.0	56	12.5	11	0.0	5	0.0	37	0.0	37	0.0	38	15.8	38	15.8	32	6.3	32	6.3				
P4	72	1.4	101	0.0	14	14.3	44	2.3	15	20	13	7.7	54	9.3	54	3.7	10	10.0	9	0.0	38	0.0	38	0.0	38	15.8	38	15.8	32	6.3	32	6.3				
M1	73	4.1	97	7.2	13	38.5	46	19.6	14	21.4	14	14.3	52	23.1	51	33.3	9	44.4	8	25.0	51	7.8	51	7.8	42	19.0	42	19.0	42	19.0	42	19.0				
M2	75	2.7	111	0.9	14	28.6	48	10.4	12	16.7	12	0	48	39.6	53	20.8	11	45.5	8	12.5	43	16.3	43	16.3	24	25.0	24	25.0	19	10.5	19	10.5				
M3	54	9.3	98	6.1	15	26.7	28	7.1	9	44.4	12	25	28	42.9	42	28.6	7	42.9	7	14.3	24	25.0	24	25.0	19	10.5	19	10.5	19	10.5	19	10.5				
Total	523	2.5	783	2.0	126	16.7	271	8.1	109	19.3	99	8.0	389	19.8	417	14.4	85	15.3	58	6.9	290	7.9	290	7.9	241	7.5	241	7.5	241	7.5	241	7.5				
Mandible	52	0.0	90	1.1	11	0.0	22	0.0	13	0	11	0	45	0.0	60	0.0	5	0.0	3	0.0	27	0.0	27	0.0	12	0.0	12	0.0	14	0.0	14	0.0				
I1	67	0.0	111	0.9	12	0.0	22	0.0	15	0	14	14.3	54	1.9	72	1.4	7	0.0	5	0.0	32	0.0	32	0.0	14	0.0	14	0.0	14	0.0	14	0.0				
I2	68	0.0	120	0.0	12	0.0	26	7.7	15	0	16	12.5	58	5.2	71	2.8	11	9.1	7	0.0	34	0.0	34	0.0	18	0.0	18	0.0	18	0.0	18	0.0				
C	77	1.3	123	0.8	12	8.3	27	0.0	17	11.8	16	18.6	56	7.1	70	8.6	10	10.0	5	0.0	33	0.0	33	0.0	20	0.0	20	0.0	20	0.0	20	0.0				
P3	82	0.0	137	0.0	13	0.0	27	0.0	16	6.3	17	5.9	51	15.7	69	7.2	11	0.0	5	0.0	32	0.0	32	0.0	20	0.0	20	0.0	20	0.0	20	0.0				
P4	83	3.6	154	1.9	13	7.7	32	3.2	16	0	18	27.8	45	28.9	62	17.7	9	22.2	5	0.0	35	11.4	35	11.4	28	14.3	28	14.3	28	14.3	28	14.3				
M1	91	9.9	149	2.7	13	15.4	28	0.0	15	0	15	20	54	16.7	64	21.9	7	28.6	7	14.3	26	11.5	26	11.5	27	3.7	27	3.7	27	3.7	27	3.7				
M2	59	22.0	124	4.8	14	28.6	27	11.1	17	23.5	13	38.5	32	18.8	49	16.3	5	40.0	6	16.7	21	14.3	21	14.3	24	12.5	24	12.5	24	12.5	24	12.5				
M3	579	4.5	1008	1.6	100	8.0	211	2.8	124	5.6	120	17.5	395	11.1	517	9.1	65	12.3	43	4.7	240	4.2	240	4.2	163	4.9	163	4.9	163	4.9	163	4.9				
Total	110	3.5	226	1.8	226	12.8	482	5.8	233	12.0	219	13.2	784	15.4	934	11.5	150	14.0	101	5.9	530	6.2	530	6.2	404	6.4	404	6.4	404	6.4	404	6.4				

^a Molnar 1971b.

^b Hoffman 1987.

^c Walker and Erlandson 1986.

TABLE 5. Carious Lesions in Adult Molars by Age Group (N, %)

Tooth ^a	Age-at-death							Total
	21-25	26-30	31-35	36-40	41-45	46-50	51-55	
UM1	20 (0.0)	21 (4.7)	34 (8.8)	52 (15.4)	52 (3.8)	48 (6.2)	8 (0.0)	235 (7.7)
UM2	19 (0.0)	21 (9.5)	29 (3.4)	55 (3.6)	52 (1.9)	62 (0.0)	8 (0.0)	246 (2.4)
UM3	10 (0.0)	15 (13.3)	22 (9.1)	41 (7.3)	44 (2.3)	56 (12.5)	5 (20.0)	193 (8.3)
LM1	24 (0.0)	28 (0.0)	37 (5.4)	59 (0.0)	81 (0.0)	80 (5.0)	12 (0.0)	321 (1.8)
LM2	24 (16.7)	31 (3.2)	35 (11.4)	62 (1.6)	79 (3.8)	78 (2.6)	11 (9.1)	320 (5.1)
LM3	20 (15.0)	25 (8.0)	21 (23.8)	44 (15.9)	72 (9.7)	54 (3.7)	8 (25.0)	244 (11.5)
Total	117 (6.0)	141 (5.7)	178 (9.5)	313 (7.0)	380 (3.7)	378 (4.8)	52 (7.7)	1559 (5.8)

^a Antimeres are combined.

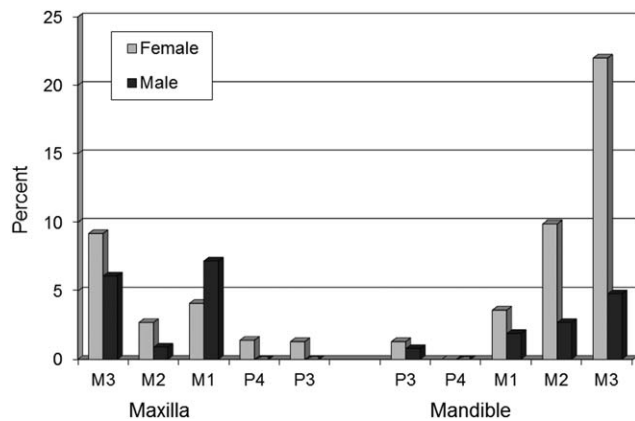


Fig. 3. Frequencies of carious lesions in maxillary and mandibular teeth of females and males.

dynamic interplay between unique features of subgingival microenvironments and cultural factors. Examination of periodontal recession levels for each tooth position (Fig. 7) shows a population with very moderate amounts of alveolar bone resorption. Considering the upper and lower dentitions separately, the upper dentition shows 59.9% ($n = 1001/1671$) of the teeth with CEJ to AC measurements greater than 2 mm and 27.2% ($n = 454/1671$) greater than 3 mm. The lower dentition exhibits 76.9% ($n = 1657/2154$) of the teeth with CEJ to AC measurements greater than 2 mm and 56.3% ($n = 1213/2154$) greater than 3 mm. Females in this sample generally have greater amounts of periodontal recession than males, however the difference in frequencies is not statistically significant ($\chi^2 = 0.120$, P -value = 0.729). The overall frequency of individuals that exhibit periodontal disease (CEJ to AC measurements greater than 2 mm and evidence of alveolar bone porosity) is 17.8%.

Examination of periodontal disease frequencies reported in Table 6 shows that unlike most other population samples, females in the CA-CCO-548 sample have overall frequencies of periodontal disease that are higher than males (20.3% vs. 16.6%). The difference is statistically significant ($\chi^2 = 10.41$, P -value = 0.001). Examination of the frequencies of periapical abscesses reported in Table 6 reveals a relatively high number of lesions across the entire sample, especially in the region of the maxillary and mandibular first and second molars. The overall frequency of periapical abscesses is 10.7% ($n = 409/3825$). Females show a higher frequency of periapical abscesses than do males (14.0% vs. 10.8%). The

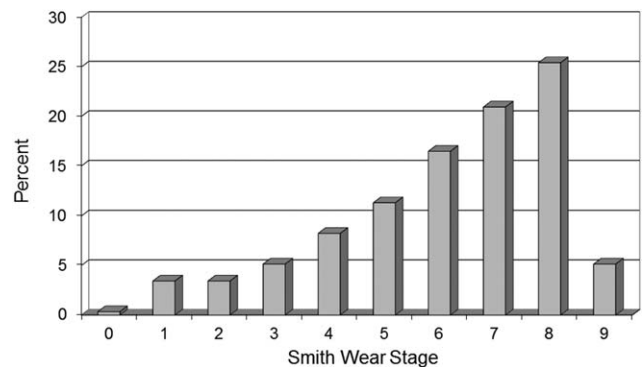


Fig. 4. Frequencies of Smith (1984) composite dental wear scores for CA-CCO-548.

difference is statistically significant ($\chi^2 = 7.92$, P -value = 0.0049).

Taken together, the frequency and pattern of periodontal recession and periapical abscesses observed in this population is one that cannot be fully explained by disease. That is, while the rates of periodontal recession in excess of 2 mm are high at 69.5% ($n = 2658/3825$), less than one-third of these tooth positions exhibit signs of periodontal disease. In fact, the teeth with the highest overall rates of recession (lower I1-P4) have some of the lowest frequencies of periodontal disease. The most likely explanation for the levels of recession in these teeth is super-eruption due to excessive wear. Super-eruption of the dentition (continuous eruption) has been suggested as a physiological compensatory mechanism for horizontal enamel loss in humans (Newman, 1999; Haytac et al., 2013). Continuous eruption of this type increases the distance between the CEJ and the AC (Newman, 1999). Continuous tooth eruption has been documented in populations that exhibit excessive dental wear (Levers and Darling, 1983; Whittaker et al., 1985; Varrela et al., 1995). While continuous eruption has been shown to occur in humans irrespective of wear, it does seem to be correlated with the availability of inter-occlusal space (Levers and Darling, 1983; Newman, 1999).

DISCUSSION

Four notable features emerge from the statistical analyses of the CA-CCO-548 dentitions. The sample exhibits 1) extraordinarily low frequencies of carious lesions, 2) very low frequencies of periodontal disease, 3) high

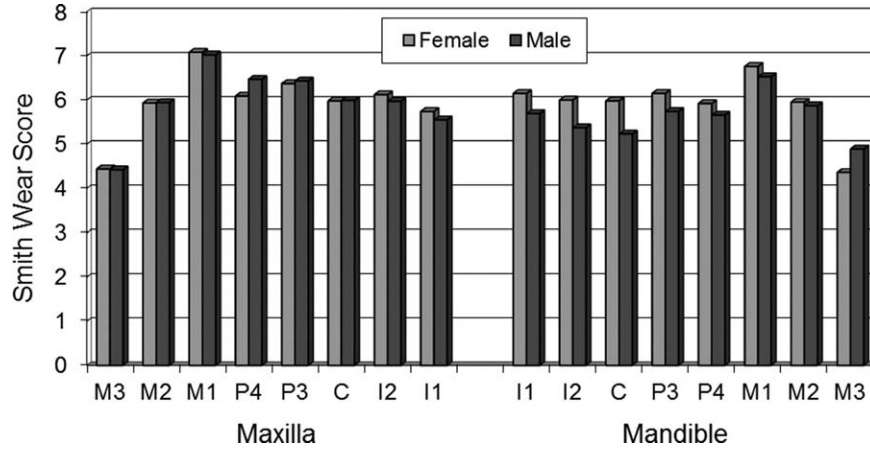


Fig. 5. Mean dental wear by tooth class and sex.

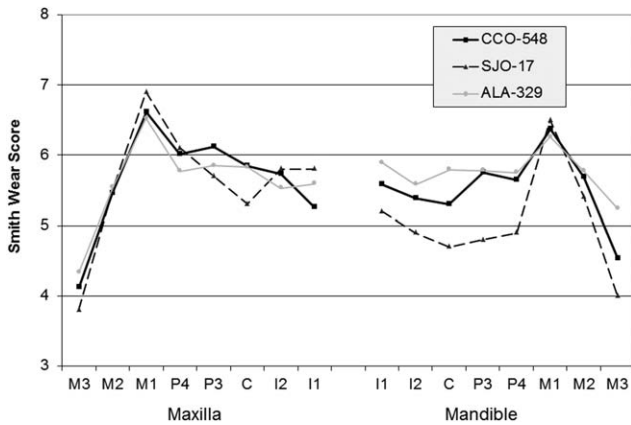


Fig. 6. Mean dental wear by tooth class and sex for CA-CCO-548 compared to two other sites, CA-SJO-17 and CA-ALA-329 (comparative data from Molnar 1971 and Jurmain 1990).

frequencies of periapical abscesses, 4) and remarkably high levels of dental wear. Because the etiologies of the pathological conditions are interrelated, it is logical to examine the relationship and potential influence of each condition on the others. For instance, both caries and the periodontal diseases have as their primary etiological agent a shift of the oral microflora from commensal to facultative pathogenic in the diseased individual (Marsh and Devine, 2011; Takahashi and Nyvad, 2011; Dentino et al., 2013). Another key shared risk contributor for the pathologic processes is poor oral hygiene (Stabholz et al., 2010; Rodrigues et al., 2011; Dentino et al., 2013). Dental wear also has a potential influence on all three of the pathologic processes (Newman, 1999; Caglar et al., 2007; Buzon and Bombak, 2010).

Dental caries has been used in bioarchaeological studies as an indicator of overall dental health and dietary changes in past populations (Larsen et al., 1991; Sciulli, 1997; Temple and Larsen, 2007; Slaus et al., 2011; Williams and Murphy, 2013). Much of the literature pertaining to the influence of dietary change on overall health has focused on the adoption of agriculture (Larsen et al., 1991; Lukacs, 1992; Temple and Larsen, 2007; Watson, 2008; Cucina et al., 2011; Halcrow et al., 2013). The conclusion of this body of research is that the increase in carbohydrate consumption with the transition

to agriculture precipitates an increase in caries prevalence. Clinical research indicates that the relationship between complex carbohydrates and caries is more complicated than much of the bioarchaeological research suggests (Lingström et al., 1994; Beighton et al., 1996; Lingström et al., 2000; Hujoel, 2009; Moynihan, 2012).

Clinical investigation has found that the relationship between consumption of low molecular weight (LMW) carbohydrates (white flour and white sugar) and caries is rather direct (Lingström et al., 2000; Moynihan and Kelly, 2014). In addition to being fermentable, LMW carbohydrates such as sucrose act as a substrate for the synthesis of polysaccharides in dental plaque (Bowen, 2002). Long-term consumption of LMW carbohydrates therefore lowers oral pH and shifts the balance of microflora from predominantly commensal to more cariogenic (Paes Leme et al., 2006; Parisotto et al., 2010). Nonhuman research has found a similar synergistic relationship between LMW carbohydrates and subgingival pathogenic bacteria associated with periodontal disease (Aguirre et al., 2012; Seneviratne et al., 2013). The relationship between complex carbohydrate consumption and caries rates is far less conclusive (Lingström et al., 2000). In fact, some research has shown that the consumption of starches enhances oral calcification of tooth enamel (Hidaka and Oishi, 2007). Other research has shown that starch encourages the proliferation of commensal bacteria found in the oral cavity at the potential expense of pathogenic species (Nikitkova et al., 2012).

In vitro biofilm and nonhuman caries studies have shown that complex carbohydrates are substantially less cariogenic than sucrose or glucose (Duarte et al., 2008; Thurnheer et al., 2008). *In vivo* human research has also shown that complex carbohydrates that have not undergone extensive processing generally do not offer sufficient bioavailability of the sugars necessary for cariogenic bacterial proliferation (Lingström et al., 2000) or do so at a significantly reduced rate (Moynihan, 2012). Research has shown that the manner in which the carbohydrate is processed is far more important than the amount or type of the carbohydrate consumed (Grenby, 1990; Harper et al., 1985). Modern commercial processing methods used for complex carbohydrates, such as high-temperature extrusion, explosion puffing, or “instantization,” render starches with far higher glycemic and hydrolytic indices than their non-commercially processed counterparts (Brand et al., 1985; Ross et al.,

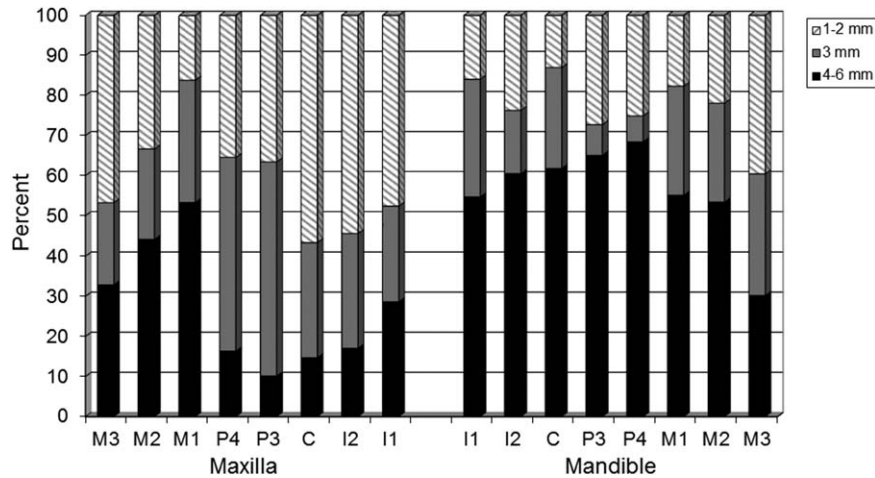


Fig. 7. Mean periodontal recession by tooth class for CA-CCO-548.

TABLE 6. Frequencies of periodontal disease (PD) and periapical abscesses (AA) in maxillary and mandibular dentitions

	Total sample			Female			Male		
	N	% PD	% AA	N	%PD	%AA	N	%PD	%AA
Maxilla									
I1	185	14.6	9.7	52	17.3	11.5	101	12.9	9.9
I2	193	9.8	6.7	59	13.6	8.5	102	6.9	5.9
C	224	13.8	8.9	69	15.9	10.1	110	11.8	8.2
P3	227	7.5	8.4	67	7.5	8.9	106	5.7	8.5
P4	226	11.9	9.3	67	23.9	16.4	106	9.4	8.5
M1	236	44.9	34.7	71	43.7	40.8	109	41.3	38.5
M2	213	27.2	12.2	60	26.7	18.3	105	19.1	13.3
M3	167	11.4	5.4	43	4.6	2.3	89	13.5	9.0
Total	1671	18.2	12.4	488	20.1	15.6	828	15.2	12.9
Mandible									
I1	201	4.5	1.5	61	8.2	3.3	96	4.2	1.0
I2	228	6.6	3.5	73	11.0	9.6	106	2.8	0.9
C	254	11.4	5.5	77	18.2	13.0	115	5.2	0.9
P3	275	10.2	6.2	88	19.3	12.5	121	1.7	1.7
P4	295	8.1	5.1	88	11.4	6.8	135	5.9	5.2
M1	323	41.8	23.8	91	52.8	30.8	150	46.0	24.7
M2	320	31.3	15.3	91	25.3	15.4	148	41.2	18.9
M3	258	14.3	7.4	71	8.5	5.6	126	18.3	10.3
Total	2154	17.5	9.4	640	20.5	12.8	997	17.7	9.0
Site total	3825	17.8	10.7	1128	20.3	14.0	1825	16.6	10.8

1987). In other words, starches that have been gelatinized and partially degraded during processing are far more cariogenic than starches processed in other manners (Lingström et al., 2000). Another important finding is that the combination of sucrose and starch vastly amplifies the cariogenic potential of sucrose alone (Campain et al., 2003; Duarte et al., 2008; Moynihan, 2012). The observations concerning the cariogenic propensity of complex carbohydrates versus LMW carbohydrates from current clinical research highlight an important finding for bioarchaeological research. That is, the cariogenic capacity of complex carbohydrates found in the modern diet is a direct result of the means and methods of manufacture and not an inherent characteristic of the food items found in preindustrial diets. Analysis of microbiota in dental calculus for populations spanning the last 10,000 years confirms this association (Adler et al., 2013). That is, shifts in the microbiota from predominantly commensal to predominantly pathogenic did not occur until the Industrial Revolution.

Dietary analysis for CA-CCO-548 provides important evidence for the cariogenic potential of the subsistence regimen at this site. Isotopic analysis for CA-CCO-548 shows that the majority of the diet derived from terrestrial and freshwater resources with a very small contribution from marine resources (Bartelink et al., 2010). Despite the close proximity of CA-CCO-548 to San Francisco Bay, the abundance of local terrestrial and fresh water aquatic resources encouraged the exploitation of resources predominantly from the immediate vicinity rather than the more distant marine resources. Analysis of stable isotopes and paleobotanical evidence from CA-CCO-548 indicates the substantial inclusion of acorns in the diet (Bartelink et al., 2010; Wohlgemuth, 2010). Acorns have a very high carbohydrate content (Soni et al., 1993; Correia et al., 2009; Yoo et al., 2012) with an amylose content similar to apples (Stevenson et al., 2006). The amylose and amylopectin content of acorns is considerably higher than that of corn, wheat, rice, and all other A-type starches (Soni et al., 1993; Stevenson

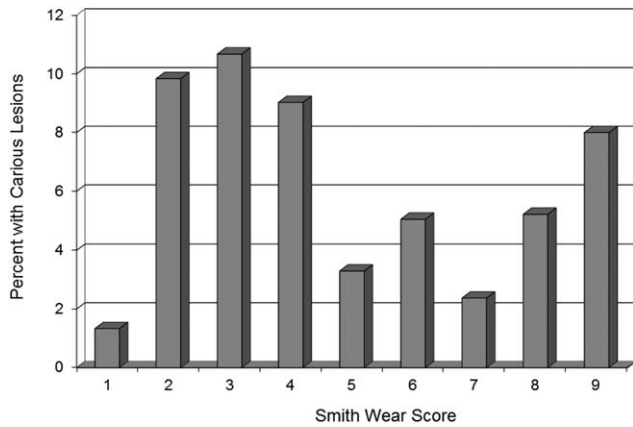


Fig. 8. Frequency of carious lesions for each wear score in the molar series.

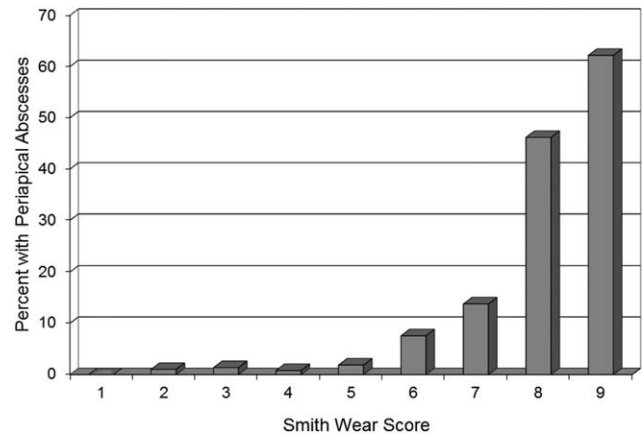


Fig. 9. Frequency of periapical lesions for each wear score in the molar series.

et al., 2006; Yoo et al., 2012). The high tannin content of acorns necessitated an intensive processing and cooking procedure which approximated some of the techniques of modern LMW starch production (Basgall, 1987; Margolin, 1989; Jacknis, 2004). For instance, the repeated grinding, sifting, and drying involved in the traditional preparation of acorns is similar to modern techniques of mechanical reduction which make starches highly susceptible to gelatinization (Lingström et al., 2000). In addition, acorns have a very low temperature of gelatinization (Stevenson et al., 2006) compared with other starches (Ashogbon and Akintayo, 2013). High water-starch ratios significantly decrease the temperature at which gelatinization occurs (Lingström et al., 2000). Water-leaching to remove tannins and hot water meal cooking techniques contributed to high water-starch ratios in traditional acorn preparation (Basgall, 1987; Margolin, 1989; Jacknis, 2004). Considering the high amylose and amylopectin content of acorns, the intensive processing techniques associated with acorn consumption, and the demonstrated relationship between a high carbohydrate diet and prevalence of caries, one would expect a high prevalence of caries in the CA-CCO-548 sample. However, not only is caries prevalence very low at CA-CCO-548, it is one of the lowest recorded in pre-contact Central California. Recent clinical research regarding the etiologies of oral pathologies offers possible explanations for the low prevalence of caries in a population consuming a carbohydrate-rich diet.

One possible explanation for the exceptionally low frequencies of carious lesions in this sample is excessive dental wear. The results reported here for CA-CCO-548 show an inverse relationship between wear and frequencies of carious lesions. That is, while the overall frequency of carious lesions is remarkably low at 2.5%, the overall average Smith wear score is notably high at 6.1 out of nine. Isolating the comparison to only the molars, the teeth most susceptible to caries, the relationship is statistically significant ($\chi^2 = 30.301$, P -value = 0.000). However, the relationship between dental wear and carious lesions is only weakly correlated (Tau-b = -0.052). Examination of the distribution of carious lesion frequencies for each wear score reveals why the relationship is very weakly correlated (Fig. 8). While the inverse relationship does generally hold true for comparison of wear scores 2 through 7, teeth with the wear scores of 1, 8, or 9 do not follow the overall trend. Results of this

analysis do not provide strong support for the cariostatic influence of dental wear.

While a causative relationship between degree of wear and prevalence of caries is not supported here, a relationship between dental wear and periapical lesions is supported. Examination of dental wear scores for each tooth position and the corresponding frequency of periapical lesions for that position show a positive correlation. Isolating the comparison with the molars, the teeth most susceptible to periapical lesions, low wear scores correspond to low frequencies of lesions (Fig. 9). Likewise, as the wear progresses, frequencies of periapical abscesses increase. The relationship is statistically significant ($\chi^2 = 378.88$, P -value = 0.000) and relatively strong (Tau-b = 0.386). Researchers have noted the likely interaction between excessive dental wear and frequency of periapical lesions (Clarke et al., 1986; Jurmain, 1990; Clarke and Hirsch, 1991). Excessive dental wear causes compressive injury of the dental pulp which releases inflammatory mediators leading to inflammation of the periodontal ligament (Torobinejad and Bakland, 1980). Clinical research has demonstrated a direct relationship between physiological compressive force on the periodontal ligament and the stimulation of inflammation related genes (El-Awady et al., 2013). Bioarchaeological research has shown that even in the relative absence of other evidence of periodontal disease, populations that show excessive wear also exhibit high frequencies of periapical abscesses (Clarke et al., 1986; Hall et al., 1986; Clarke and Hirsch, 1991). Statistical analysis in this population sample shows a significant and strong positive correlation between severe wear and periapical lesion frequency.

Another possibility connecting the low frequencies of carious lesions to the high levels of attrition is revealed by examining not only the extent of wear but also the specific patterns of wear. In particular, when the corresponding maxillary and mandibular dentitions are articulated in anatomical position, a revealing pattern of wear emerges. In the majority of intact dentitions from this site, the maxillary and mandibular teeth are not capable of direct occlusion due to the wear facets that have developed (Fig. 10). The pattern of wear has been shown to most likely result from non-alimentary behavior such as basket-making or cordage manufacture (Grant, 2010). Dietary items and non-alimentary



Fig. 10. Examples of non-alimentary dental wear in CA-CCO-548 (photos by Dave Grant).

material consistently in contact with the oral cavity have the capacity to alter the microbial environment, potentially disrupting the ability of pathogenic bacteria to displace the commensal bacteria. Specifically, phenolic compounds found in dietary and non-dietary plant materials have antimicrobial properties (Lattanzio et al., 2008; Dai and Mumper, 2010).

Many naturally occurring phenolic plant compounds have antimicrobial properties specific to human pathogens (Chung et al., 1998; Puupponen-Pimiä et al., 2005; Friedman, 2007). Salicylic acid is a naturally occurring phenolic plant hormone that has been used medicinally for more than 2,000 years (Vlot et al., 2009). The primary natural source of salicylic acid is the willow tree (*Salix sp.*). Willow was a ubiquitous material used in Central California Native American basketry, as well as a wide range of household applications including the manufacture of bows, arrows, women's skirts, mats, cordage, and the construction of dwellings (Elsasser, 1978; Farmer, 2010; Shanks and Shanks, 2010). While basket making was largely confined to females, construction of other household items using willow was not exclusively a female occupation. The traditional method of fiber and willow bark preparation (still practiced modernly) is via oral modification and manipulation of the materials (Merriam, 1967; Newman, 1974; Bocek, 1984; Shanks and Shanks, 2006). In plants, salicylic acid plays critical roles in development and metabolism (Vlot et al., 2009). It also serves a key function in plant disease resistance by inducing the production of pathogenesis-related proteins (Kawano and Furuichi, 2007). These proteins offer resistance to viral, bacterial, and fungal pathogens. When used medicinally in humans and *in vitro* experiments, salicylic acid has been found to inhibit the bacterial colonization of pathogenic organisms including species of *Staphylococcus*, *Streptococcus*, *Porphyromonas*,

and *Salmonella* (Kupferwasser et al., 2003; Bandara et al., 2004; Guinta et al., 2011; Rivero-Cruz et al., 2011). Although the specific mechanism of bacterial inhibition is not fully understood, the effect has been documented in a wide variety of contexts including specific intra-oral inhibition of *Streptococcus mutans* (Muroi and Kubo, 1993).

Willow bark would have been introduced to the oral cavity on a daily basis through the manufacture of basketry and other household materials. Because willow baskets were used in the preparation and boiling of acorns (Mayer, 1976; Bibby, 2012), salicylic acid would have been inadvertently added to the diet via this route as well. Another phenolic compound introduced into the oral cavity was tannic acid. Paleobotanical evidence from CA-CCO-548 shows a substantial reliance on acorns as well as other nuts (e.g., buckeye) with a tannic acid component (Wohlgemuth, 2010). Tannic acid, a compound found in significant quantities in acorns, has been shown to inhibit the growth of many pathogenic bacteria including *Streptococcus mutans* (Chung et al., 1993; Serano et al., 2009). While the traditional preparation methods extracted most of the tannic acids before consumption (Gifford, 1936; Mayer, 1976), some of the acid remained after processing. The combined effect of daily introduction of these phenolic plant substances into the oral cavity inevitably would be the lifelong suppression of pathogenic oral bacteria.

The pattern of periodontal pathology observed in the CA-CCO-548 sample is also consistent with the interpretation of oral microbial suppression. Comparison of periodontal disease frequencies between archaeological populations is difficult because of the general lack of standardization in data collection. Studies from archaeological populations do exist (Clarke et al., 1986; Delgado-Darias, 2006; Eshed et al., 2006; Klaus and Tam, 2010;

DeWitte and Bekvalac, 2011; Wasterlain et al., 2011; DeWitte, 2012; Vodanović et al., 2012) as does abundant data from modern populations (e.g., Baelum and López, 2013; Genco and Borgnakke, 2013; Haytac et al., 2013). In all available comparisons, the frequencies of periodontal disease reported for the CA-CCO-548 sample are exceptionally low. While periodontal disease has a multifactorial etiology, the primary agents are facultative pathogenic bacterial species, which if permitted to proliferate will initiate disease (Dentino et al., 2013). Suppression of these pathogenic bacteria is the key to preventing or controlling the disease process (Dentino et al., 2013). The primary etiological agents associated with periapical abscesses are also facultative pathogenic bacterial species (Siqueira and Rôças, 2009). Because the infection originates in the root canal system (Siqueira and Rôças, 2009), clinical treatment of periapical lesions requires mechanical debridement to gain access to the infected endodontic space followed by systemic antibiotic treatment (Dahlén, 2002). Therefore, pathogenic bacteria that proliferate in endodontic spaces are protected from antibacterial agents in the oral cavity. The bacterial suppression effect that would apply to caries and periodontitis therefore would not apply in the same way to risk of periapical lesions.

CONCLUSIONS

The results of this study show a population sample that exhibits extraordinarily low frequencies of carious lesions and periodontitis, high frequencies of periapical lesions, and remarkably high levels of dental wear. Wear was examined as a causative agent for the low prevalence of caries; however, statistical analysis shows a very weak association between the two. Statistical analysis does show a significant and strong correlation between wear and prevalence of periapical lesions. A more cogent explanation for the low prevalence of caries and periodontitis is provided by examination of the likely cause of the excessive dental wear rather than its extent. The individuals that comprise the CA-CCO-548 population sample exhibit an unusual wear pattern that is the likely result of non-dietary abrasion, specifically the manufacture of basketry. Materials used for Native Californian basketry contain naturally high levels of antimicrobial compounds. In addition, staple constituents of the documented dietary regime for this population also contain high levels of antimicrobial compounds. At least two of these antimicrobial compounds, salicylic and tannic acids, have been clinically documented to inhibit the primary bacterial species implicated in dental caries and periodontal disease.

This research offers a novel explanation for the remarkably low frequencies of dental caries that have been noted by virtually every bioarchaeological study of California Native American populations. The evidence presented here suggests an explanation integrating the current etiological explanations for infections of the oral cavity from the clinical literature, information from the ethnographic record pertaining to subsistence and activity patterns in Native Californian populations, and statistical analysis of specific disease and wear patterns in a large and temporally well-defined population sample. This study shows a correlation between lowered prevalence of caries and periodontitis, elevated level of dental wear and prevalence of periapical lesions, and cultural

practices that introduced antibacterial compounds into the oral cavity.

ACKNOWLEDGMENTS

The author thanks Holman and Associates Archaeological Consultants and Shea Homes for the opportunity to work with the human skeletal remains from this site. He also thanks Ms. Ramona Garibay, site monitor and Most Likely Descendant who offered me the opportunity to work with her ancestors. He thanks Ms. Garibay and Ms. Kathy Wallace who both provided invaluable insight into traditional Miwok and Ohlone lifeways. Thanks also to Patricia Lambert, Judi Strebel, Cynthia Wilczak, and two anonymous reviewers whose comments and corrections greatly improved this work.

LITERATURE CITED

- Abbott PV. 2004. Classification, diagnosis and clinical manifestations of apical periodontitis. *Endod Topics* 8:36–54.
- Adler CJ, Dobney K, Weyrich LS, Kaidonis J, Walker AW, Haak W, Bradshaw CJA, Townsend G, Soltysiak A, Alt KW, Parkhill J, Cooper A. 2013. Sequencing ancient calcified dental plaque shows changes in oral microbiota with dietary shifts of the Neolithic and Industrial revolutions. *Nat Genet* 45:450–455.
- Afroughi S, Faghihzadeh S, Khaledi MJ, Motlagh MG. 2010. Dental caries analysis in 3–5 years old children: a spatial modeling. *Arch Oral Biol* 55:374–378.
- Aguirre JI, Akhter MP, Kimmel DB, Pingel J, Xia X, Williams A, Jorgensen M, Edmonds K, Lee JY, Reinhard MK, Battles AH, Kesavalu L, Wronski TJ. 2012. Enhanced alveolar bone loss in a model of non-invasive periodontitis in rice rats. *Oral Dis* 18:459–468.
- Ashogbon AO, Akintayo ET. 2013. Recent trend in the physical and chemical modification of starches from different botanical sources: a review. *Starch-Stärke* 65:1–17.
- Baelum V, López R. 2013. Periodontal disease epidemiology—learned and unlearned? *Periodontol* 2000 62:37–58.
- Bandara BMK, Sankaridurg PR, Wilcox MDP. 2004. Non-steroidal anti inflammatory agents decrease bacterial colonisation of contact lenses and prevent adhesion to human corneal epithelial cells. *Curr Eye Res* 29:245–251.
- Bartelink EJ. 2006. Resource Intensification in Pre-Contact Central California: a Bioarchaeological Perspective on Diet and Health Patterns Among Hunter-Gatherers from the Lower Sacramento Valley and San Francisco Bay. PhD Dissertation, Department of Anthropology, Texas A&M University, College Station, Texas.
- Bartelink EJ, Beasley MM, Eerkens J, Gardner KS, Jorgenson G. 2010. Paleodietary analysis of human burials: stable carbon and nitrogen isotope results. In: Wiberg R, editor. *Archaeological investigations at CA-CCO-18/548: Final report for the Vineyards at Marsh Creek Project, Contra Costa County, California*. San Francisco: Holman and Associates Archaeological Consultants. p 409–419.
- Basgall ME. 1987. Resource intensification among hunter-gatherers: acorn economies in prehistoric California. *Res Econ Anthropol* 9:21–52.
- Beaton JM. 1991. Extensification and intensification in central California prehistory. *Antiquity* 65:946–952.
- Beighton D, Adamson A, Rugg-Gunn A. 1996. Associations between dietary intake, dental caries experience and salivary bacterial levels in 12-year-old English schoolchildren. *Arch Oral Biol* 41:271–280.
- Bibby B. 2012. *Essential art: native basketry from the California Indian Heritage Center*. Berkeley, CA: Heyday.
- Blake J. 2011. *Nonalimentary tooth use in ancient California*. Master's Thesis. San Francisco State University, CA: Department of Anthropology.

- Bocek BR. 1984. Ethnobotany of Costanoan Indians, California, based on the collections by John P. Harrington. *Econ Bot* 38: 240–255.
- Bowen WH. 2002. Do we need to be concerned about dental caries in the coming millennium? *Crit Rev Oral Biol Med* 13: 126–131.
- Brand IC, Nicholson PL, Thorburn AW, Truswell AS. 1985. Food processing and the glycemic index. *Am J Clin Nutr* 42: 1192–1196.
- Brauner AW, Conrads G. 1995. Studies into the microbial spectrum of apical periodontitis. *Int Endod J* 28:244–248.
- Buduneli N, Kinane DF. 2011. Host-derived diagnostic markers related to soft tissue destruction and bone degradation in periodontitis. *J Clin Periodontol* 38:85–105.
- Buikstra JE, Ubelaker DH. 1994. Standards for Data Collection from Human Skeletal Remains. Arkansas Archaeological Survey Report No. 44. Fayetteville, AR.
- Buzon MR, Bombak A. 2010. Dental disease in the Nile Valley during the New Kingdom. *Int J Osteoarchaeol* 20:371–387.
- Caglar E, Kusu OO, Sandalli N, Ari I. 2007. Prevalence of dental caries and tooth wear in a Byzantine population (13th c. A.D.) from northwest Turkey. *Arch Oral Biol* 52:1136–1145.
- Campaign AC, Morgan MV, Evans RW, Ugoni A, Adams GG, Conn JA, Watson MJ. 2003. Sugar-starch combinations in food and the relationship to dental caries in low-risk adolescents. *Eur J Oral Sci* 111:316–325.
- Caufield PW, Cutter GR, Dasanayake AP. 1993. Initial acquisition of mutans streptococci by infants: evidence for a discrete window of infectivity. *J Dent Res* 72:37–45.
- Cekici A, Kantarci A, Haturk H, Van Dyke TE. 2014. Inflammatory and immune pathways in the pathogenesis of periodontal disease. *Periodontol* 2000 64:57–80.
- Chung KT, Lu Z, Chou MW. 1998. Mechanism of inhibition of tannic acid and related compounds on the growth of intestinal bacteria. *Food Chem Toxicol* 36:1053–1060.
- Chung KT, Stevens SE, Lin WF, Wei CI. 1993. Growth inhibition of selected food-borne bacteria by tannic acid, propyl galate and related compounds. *Lett Appl Microbiol* 17:29–32.
- Clarke NG, Carey SE, Srikandi W, Hirsch RS, Leppard PI. 1986. Periodontal disease in ancient populations. *Am J Phys Anthropol* 71:173–183.
- Clarke NG, Hirsch RS. 1991. Tooth dislocation: the relationship with tooth wear and dental abscesses. *Am J Phys Anthropol* 85:293–298.
- Corcuera MT, Gómez-Lus ML, Gómez-Aguado F, Maestre JR, del Carmen Ramos M, Alonso MJ, Prieto J. 2013. Morphological plasticity of *Streptococcus oralis* isolates for biofilm production, invasiveness, and architectural patterns. *Arch Oral Biol* 58:1584–1593.
- Correia PR, Leitão AE, Beirão-da-Costa ML. 2009. Effect of drying temperatures on chemical and morphological properties of acorn flours. *Int J Food Sci Technol* 44:1729–1736.
- Costerton JW. 1999. Introduction to biofilm. *Int J Antimicrob Agents* 11:217–221.
- Coykendall AL. 1989. Classification and identification of the viridans streptococci. *Clin Microbiol Rev* 2:315–328.
- Cucina A, Cantillo CP, Sosa TS, Tiesler V. 2011. Carious lesions and maize consumption among the prehispanic Maya: an analysis of a coastal community in northern Yucatan. *Am J Phys Anthropol* 145:560–567.
- Dahlén, G. 2002. Microbiology and treatment of dental abscesses and periodontal-endodontic lesions. *Periodontol* 2000 28:206–239.
- Dai J, Mumper RS. 2010. Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules* 15:7313–7352.
- Davies DM, Picton DCA, Alexander AG. 1969. An objective method of assessing periodontal condition in human skulls. *J Periodont Res* 4:74–77.
- Davies GM, Worthington HV, Clarkson JE, Thomas P, Davies RM. 2001. The use of fibre-optic transillumination in general dental practice. *Br Dent J* 191:145–147.
- Delgado-Darias T, Velasco-Vázquez J, Arnay-de-la-Rosa M, Martín-Rodríguez E, González-Reimers E. 2006. Calculus, periodontal disease and tooth decay among the prehispanic population from Gran Canaria. *J Archaeol Sci* 33:663–670.
- Dentino A, Lee S, Mailhot J, Hefti AF. 2013. Principles of periodontology. *Periodontol* 2000 61:16–53.
- Deter CA. 2009. Gradients of occlusal wear in hunter-gatherers and agriculturalists. *Am J Phys Anthropol* 138:247–254.
- DeWitte SN. 2012. Sex differences in periodontal disease in catastrophic and attritional assemblages from medieval London. *Am J Phys Anthropol* 149:405–416.
- DeWitte SN, Bekvalac J. 2011. The association between periodontal disease and periosteal lesions in the St. Mary Graces cemetery, London, England A.D. 1350–1538. *Am J Phys Anthropol* 146:609–618.
- Dias, G. and N. Tayles. 1997. ‘Abscess cavity’—a misnomer. *Int J Osteoarchaeol* 7:548–554.
- Dreier FG. 1994. Age at death estimates for the protohistoric Arikara using molar attrition rates: a new quantification method. *Int J Osteoarchaeol* 4:137–147.
- Duarte S, Klein MI, Aires CP, Cury JA, Bowen WH, Koo H. 2008. Influences of starch and sucrose on *Streptococcus mutans* biofilms. *Oral Microbiol Immunol* 23:206–212.
- Duyar İ, Erdal YS. 2003. A new approach for calibrating dental caries frequency of skeletal remains. *Homo* 54:57–70.
- Eerkens JW, Bartelink EJ. 2013. Sex-biased weaning and early childhood diet among middle Holocene hunter-gatherers in central California. *Am J Phys Anthropol* 152:471–483.
- El-Awady AR, Lapp CA, Gamal AY, Sharawy MM, Wenger KH, Cutler CW, Messer RLW. 2013. Human periodontal ligament fibroblast responses to compression in chronic periodontitis. *J Clin Periodontol* 40:661–671.
- Elsasser AB. 1978. Basketry. In: Heizer RF, editor. *Handbook of North American Indians*, Vol. 8. CA. Washington, D.C.: Smithsonian Institution. p 626–641.
- Eshed V, Gopher A, Hershkovitz I. 2006. Tooth wear and dental pathology at the advent of agriculture: new evidence from the Levant. *Am J Phys Anthropol* 130:145–159.
- Farmer JF. 2010. Basketry plants used by western American Indians. Fullerton, CA: Justin Farmer Foundation.
- Featherstone JDB. 2004. The continuum of dental caries – Evidence for a dynamic disease process. *J Dent Res* 83:C39–C42.
- Ferreira Zandoná A, Santiago E, Eckert GJ, Katz BP, Pereira de Oliveira S, Capin OR, Mau M, Zero DT. 2012. The natural history of dental caries lesions: a 4-year observational study. *J Dent Res* 91:841–846.
- Forng RY, Champagne C, Simpson W, Genco CA. 2000. Environmental cues and gene expression in *Porphyromonas gingivalis* and *Actinobacillus actinomycetemcomitans*. *Oral Dis* 6: 351–365.
- Friedman M. 2007. Overview of antibacterial, antitoxin, antiviral, and antifungal activities of tea flavonoids and teas. *Mol Nutr Food Res* 51:116–134.
- Genco RJ, Borgnakke WS. 2013. Risk factors for periodontal disease. *Periodontol* 2000 62:59–94.
- Giacaman RA, Aranedo E, Padilla C. 2010. Association between biofilm-forming isolates of mutans streptococci and caries experience in adults. *Arch Oral Biol* 55:550–554.
- Gifford EW. 1936. California Indian balanophagy. In: Lowie RH, editor. *Essays in anthropology presented to Alfred L. Kroeber*. Berkeley, CA: University of California Press. p 87–98.
- Grant D. 2010. Native Americans in the San Francisco Bay Area: patterns in ancient teeth, palimpsests of behavior. MA Thesis, San Jose State University, Department of Anthropology.
- Grenby TH. 1990. Snack foods and dental caries: investigations using laboratory animals. *Br Dent J* 168:353–361.
- Griffin MC, Snyder J, Balabuszko R, Entriken K, Wiberg R. 2010. Demography, health, and regional biodiversity. In: Wiberg R, editor. *Archaeological investigations at CA-CCO-18/548: Final report for the Vineyards at Marsh Creek Project, Contra Costa County, California*. San Francisco: Holman and Associates Archaeological Consultants. p 355–408.
- Guidara A. 2012. Discriminant function analysis for sex determination using tooth size at the Vineyards Site (CA-CCO-

- 548). Master's Thesis. San Francisco State University, CA: Department of Anthropology.
- Guinta RA, Carbone LA, Rosenberg EL, Uhrich EK, Tabak M, Chikindas LM. 2011. Slow release of salicylic acid from degrading poly (anhydride ester) polymer disrupts bimodal pH and prevents biofilm formation in *Salmonella typhimurium* MAE52. In: Bailey WC, editor. *Biofilms: formation, development and properties*. NY: Nova Science Publishers. p 649–658.
- Halcrow SE, Harris NJ, Tayles N, Ikehara-Quebral R, Pietruszewski M. 2013. From the mouths of babes: dental caries in infants and children and the intensification of agriculture in mainland Southeast Asia. *Am J Phys Anthropol* 150:409–420.
- Hall RL. 1976. Functional relationships between dental attrition and the helicoidal plane. *Am J Phys Anthropol* 45:69–76.
- Hall RL, Morrow R, Clarke JH. 1986. Dental pathology of prehistoric residents of Oregon. *Am J Phys Anthropol* 69:325–334.
- Han YW, Wang X. 2013. Mobile microbionme: oral bacteria in extra-oral infections and inflammation. *J Dent Res* 92:485–491.
- Hardwick JL. 1960. The incidence and distribution of caries throughout the ages in relation to the Englishman's diet. *Br Dent J* 108:9–17.
- Harper DS, Osborn JC, Hefferren JJ, Muller TP. 1985. Dental cariogenic evaluation of foods using human plaque pH and an experimental rat-caries model. *Arch Oral Biol* 30:455–460.
- Haytac MC, Ozcelik O, Mariotti A. 2013. Periodontal disease in men. *Periodontol* 2000 61:252–265.
- Hidaka S, Oishi A. 2007. An invitro study of the effect of some dietary components on calculus formation: regulation of calcium phosphate precipitation. *Oral Dis* 13:296–302.
- Hillson S. 2001. Recording dental caries in archaeological human remains. *Int J Osteoarchaeol* 11:249–289.
- Hoffman JM. 1987. *The Descriptive Physical Anthropology of the Cardinal Site, CA-SJO-154: a Late Middle Horizon—Early Phase I Site from Stockton, California*. Colorado College Publications in Anthropology No. 12.
- Holt SC, Ebersole JL. 2005. *Porphyromonas gingivalis*, *Treponema denticola*, and *Tannerella forsythia*: The “red complex,” a prototype polybacterial pathogenic consortium in periodontitis. *Periodontol* 2000 38:72–122.
- Hujoel P. 2009. Dietary carbohydrates and dental-systemic diseases. *J Dent Res* 88:490–502.
- Innings Å, Krabbe M, Ullberg M, Herrmann B. 2005. Identification of 43 *Streptococcus* species by pyrosequencing analysis of the rnpB gene. *J Clin Microbiol* 43:5983–5991.
- Jacknis I. 2004. Toward a culinary anthropology of native California. In: Jacknis I, editor. *Food in California Indian culture*. University of California, Berkeley: Phoebe Hearst Museum of Anthropology. p 1–119.
- Jenkinson HF. 2011. Beyond the oral microbiome. *Environ Microbiol* 13:3077–3087.
- Jurmain RD. 1990. Paleoevidence of a central California prehistoric population from CA-ALA-329: dental disease. *Am J Phys Anthropol* 81:333–342.
- Kaidonis JA, Richards LC, Townsend GC, Tansley GD. 1998. Wear of human enamel: a quantitative in vitro assessment. *J Dent Res* 77:1983–1990.
- Kaifu Y, Kasai K, Townsend GC, Richards LC. 2003. Tooth wear and the “design” of the human dentition: a perspective from evolutionary medicine. *Yearb Phys Anthropol* 46:47–61.
- Kawano T, Furuichi T. 2007. Salicylic acid as a defense-related plant hormone. In: Hayat S, Ahmad A, editors. *Salicylic acid—a plant hormone*. New York: Springer Publishing. p 277–321.
- Kebschull M, Demmer RT, Papananou PN. 2010. “Gum bug, leave my heart alone!”—Epidemiologic and mechanistic evidence linking periodontal infections and atherosclerosis. *J Dent Res* 89:879–902.
- Kerr NW, Bruce MF, Cross JF. 1990. Caries experience in medi-aeval Scots. *Am J Phys Anthropol* 83:69–76.
- Kidd EAM, Fejerskov O. 2004. What constitutes dental caries? Histopathology of carious enamel and dentin related to the action of cariogenic biofilms. *J Dent Res* 83:C35–C38.
- Kieser JA, Dennison KJ, Kaidonis JA, Huang D, Herbison PGP, Tayles NG. 2003. Patterns of dental wear in the early Maori dentition. *Int J Osteoarchaeol* 11:206–217.
- Klaus HD, Tam ME. 2010. Oral health and the postcontact adaptive transition: a contextual reconstruction of diet in Mórrope, Peru. *Am J Phys Anthropol* 141:594–609.
- Kleinberg I. 2002. A mixed-bacteria ecological approach to understanding the role of the oral bacteria in dental caries causation: an alternative to *Streptococcus mutans* and the specific-plaque hypothesis. *Crit Rev Oral Biol Med* 13:108–125.
- Koo H, Falsetta ML, Klein MI. 2013. The exopolysaccharide matrix: a virulence determinant of cariogenic biofilm. *J Dent Res* 92:1065–1073.
- Kupferwasser LI, Yeaman MR, Nast CC, Kupferwasser D, Xiong Y, Palma M, Cheung AL, Bayer AS. 2003. Salicylic acid attenuates virulence in endovascular infections by targeting global regulatory pathways in *Staphylococcus aureus*. *J Clin Invest* 112:222–233.
- Laine ML, Moustakis V, Koumakis L, Potamias G, Loos BG. 2013. Modeling susceptibility to periodontitis. *J Dent Res* 92: 45–50.
- Larsen CS, Shavit R, Griffin MC. 1991. Dental caries evidence for dietary change: an archaeological context. In: Kelley M, Larsen CS, editors. *Advances in dental anthropology*. New York: Wiley-Liss, Inc. p 179–201.
- Lattanzio V, Kroon PA, Quideau S, Treutter D. 2008. Plant phenolics—Secondary metabolites with diverse functions. In: Daayf F, Lattanzio V, editors. *Recent advances in polyphenol research*, Vol. 1. Oxford: Blackwell Publishing Ltd. p 1–35.
- Law V, Seow WK, Townsend G. 2007. Factors influencing oral colonization of mutans streptococci in young children. *Aust Dent J* 52:93–100.
- Lee A, He LH, Lyons K, Swain MV. 2012. Tooth wear and wear investigations in dentistry. *J Oral Rehabil* 39:217–225.
- Leigh RW. 1928. *Dental pathology of aboriginal California*. University of California Publications in American Archaeology and Ethnology 23:399–440.
- Levers BGH, Darling AI. 1983. Continuous eruption of some adult human teeth of ancient populations. *Arch Oral Biol* 28: 401–408.
- Liljemark WF, Bloomquist C. 1996. Human oral microbial ecology and dental caries and periodontal diseases. *Crit Rev Oral Biol Med* 7:180–198.
- Lingström P, Birkhed D, Ruben J, Arends J. 1994. Effect of frequent consumption of starchy food items on enamel and dentin demineralization and on plaque pH in situ. *J Dent Res* 73: 652–660.
- Lingström P, van Houte J, Kashket S. 2000. Food starches and dental caries. *Crit Rev Oral Biol Med* 11:366–380.
- Liu, YG, Lerner UH, Teng YA. 2010. Cytokine responses against periodontal infection: protective and destructive roles. *Periodontol* 2000 52:163–206.
- Lukacs JR. 1989. Dental paleopathology: methods for reconstructing dietary patterns. In: İşcan MY, Kennedy KAR, editors. *Reconstruction of life from the skeleton*. New York: Wiley-Liss. p 261–286.
- Lukacs JR. 1992. Dental paleopathology and agricultural intensification in South Asia: new evidence. *Am J Phys Anthropol* 87:133–150.
- Lukacs JR. 1995. The ‘caries correction factor’: a new method of calibrating dental caries rates to compensate for antemortem loss of teeth. *Int J Osteoarchaeol* 5:151–156.
- Maat GJR, Van der Velde EA. 1987. The caries-attrition competition. *Int J Anthropol* 2:281–292.
- Macek MD, Beltrán-Aguilar ED, Lockwood SA, Malvitz DM. 2003. Updated comparison of the caries susceptibility of various morphological types of permanent teeth. *J Public Health Dent* 63:174–182.
- Margolin M. 1989. *The Ohlone way: Indian life in the San Francisco-Monterey Bay area*. Berkeley, CA: Heydey Books.

- Marsh PD. 1995. The role of microbiology in models of dental caries. *Adv Dent Res* 9:244–254.
- Marsh PD, Devine DA. 2011. How is the development of dental biofilms influenced by the host? *J Clin Periodontol* 38:28–35.
- Marsh PD, Martin MV. 2009. *Oral microbiology*, 5th edition. Edinburgh, UK: Churchill Livingstone.
- Marsh PD, Moter A, Devine DA. 2011. Dental plaque biofilms: Communities, conflict, and control. *Periodontol* 2000 55:16–35.
- Mayer PJ. 1976. Miwok balanophagy: implications for the cultural development of some California acorn-eaters. Berkeley, CA: Archaeological Research Facility, Department of Anthropology, University of California.
- McCarthy H. 1993. Managing oaks and the acorn crop. In: Blackburn TC, Anderson K, editors. *Before the wilderness: environmental management by Native Californians*. Menlo Park, CA: Ballena Press. p 213–228.
- Meiklejohn C, Wyman JM, Schentag CT. 1992. Caries and attrition: dependent or independent variables? *Int J Anthropol* 7: 17–22.
- Merriam CH. 1967. *Ethnological notes on central California Indian tribes*. University of California Archaeological Survey 68(3). Berkeley CA: University of California.
- Merritt J, Qi F. 2012. The mutacins of *Streptococcus mutans*: regulation and ecology. *Mol Oral Microbiol* 27:57–69.
- Meurman JH, Sanz M, Janket S. 2004. Oral health, atherosclerosis, and cardiovascular disease. *Crit Rev Oral Biol Med* 15: 403–413.
- Meyer J. 2010. Geoarchaeology. In: Wiberg R, editor. *Archaeological investigations at CA-CCO-18/548: Final report for the Vineyards at Marsh Creek Project, Contra Costa County, California*. San Francisco: Holman and Associates Archaeological Consultants. p 65–90.
- Miles AEW. 1969. The dentition of Anglo-Saxons. *Proc R Soc Med* 62:1311–1315.
- Molnar S. 1971a. Human tooth wear, tooth function and cultural variability. *Am J Phys Anthropol* 34:175–190.
- Molnar S. 1971b. Sex, age, and tooth position as factors in the production of tooth wear. *Am Antiq* 36:182–188.
- Molnar S. 1972. Tooth wear and culture: a survey of tooth function among some prehistoric populations. *Curr Anthropol* 13: 511–526.
- Molnar S, McKee JK, Molnar I. 1983. Measurements of tooth wear among Australian Aborigines: I. Serial loss of the enamel crown. *Am J Phys Anthropol* 61:51–65.
- Moratto MJ. 1984. *California archaeology*. Orlando: Academic Press.
- Moynihan, P. 2012. The role of diet in the prevention of dental diseases. In: Limeback, H, editor. *Preventive dentistry*. New York: John Wiley and Sons, Ltd. p 99–114.
- Moynihan PJ, Kelly SAM. 2014. Effect on caries of restricting sugars intake: systematic review to inform WHO guidelines. *J Dent Res* 93:8–18.
- Muroi H, Kubo I. 1993. Bactericidal activity of anacardic acids against *Streptococcus mutans* and their potentiation. *J Agric Food Chem* 41:1780–1783.
- Nair PNR. 1997. Apical periodontitis: a dynamic encounter between root canal infection and host response. *Periodontol* 2000 13:121–148.
- Nair PNR. 2003. Non-microbial etiology: Periapical cysts sustain post-treatment apical periodontitis. *Endod Topics* 6:96–113.
- Newman HN. 1999. Attrition, eruption, and the periodontium. *J Dent Res* 78:730–734.
- Newman SC. 1974. *Indian basket weaving*. Flagstaff: Northland Press.
- Nikitkova AE, Haase EM, Scannapieco FA. 2012. Effect of starch and amylase on the expression of amylase-binding protein A in *Streptococcus gordonii*. *Mol Oral Microbiol* 27:284–294.
- Oliver RC, Brown LJ. 1993. Periodontal diseases and tooth loss. *Periodontol* 2000 2:117–127.
- Paes Leme AF, Koo H, Bellato CM, Bedi G, Cury JA. 2006. The role of sucrose in cariogenic dental biofilm formation—New insight. *J Dent Res* 85:878–887.
- Palmer RJ. 2014. Composition and development of oral bacterial communities. *Periodontol* 2000 64:20–39.
- Parisotto TM, Steiner-Oliveira C, Duque C, Peres RCR, Rodrigues LKA, Nobre-dos-Santos M. 2010. Relationship among microbiological composition and presence of dental plaque, sugar exposure, social factors and different stages of early childhood caries. *Arch Oral Biol* 55:365–373.
- Paster BJ, Dewhirst FE. 2009. Molecular microbial diagnosis. *Periodontol* 2000 51:38–44.
- Powell ML. 1985. The analysis of dental wear and caries for dietary reconstruction. In: Gilbert RI, Mielke JH, editors. *The analysis of prehistoric diets*. Orlando: Academic Press. p 307–338.
- Preshaw PM, Taylor JJ. 2011. How has research into cytokine interactions and their role in driving immune responses impacted our understanding of periodontitis? *J Clin Periodontol* 38: 60–84.
- Pretty IA. 2006. Caries detection and diagnosis: novel technologies. *J Dent* 34:727–739.
- Puupponen-Pimiä R, Nohynek L, Hartmann-Schmidlin S, Kähkönen M, Heinonen M, Määttä-Riihinen K, Oksman-Caldentey KM. 2005. Berry phenolics selectively inhibit the growth of intestinal pathogens. *J Appl Microbiol* 98:991–1000.
- Reinhardt GA. 1983. Relationship between attrition and lingual tilting in human teeth. *Am J Phys Anthropol* 61:227–237.
- Reyes L, Herrera D, Kozarov E, Roldán S, Progulsk-Fox A. 2013. Periodontal bacterial invasion and infection: contribution to atherosclerotic pathology. *J Clin Periodontol* 40:S30–S50.
- Richards LC. 1984. Principal axis analysis of dental attrition data from two Australian Aboriginal populations. *Am J Phys Anthropol* 65:5–13.
- Rivero-Cruz BE, Esturau N, Sánchez-Nieto S, Romero I, Castillo-Juárez I, Rivero-Cruz JF. 2011. Isolation of the new anacardic acid 6-[16'Z-nonadecenyl]-salicylic acid and evaluation of its antimicrobial activity against *Streptococcus mutans* and *Porphyromonas gingivalis*. *Nat Prod Res* 25:1282–1287.
- Rodrigues JA, Lussi A, Seemann R, Neuhaus KW. 2011. Prevention of crown and root caries in adults. *Periodontol* 2000 55: 231–249.
- Rosenthal JS, White GG, Sutton MQ. 2007. The central valley: a view from the catbird's seat. In: Jones TL, Klar KA, editors. *California prehistory: colonization, culture, and complexity*. Lanham, MD: Alta Mira Press. p 147–163.
- Ross SW, Brand IC, Thorburn AW, Truswell AS. 1987. Glycemic index of processed wheat products. *Am J Clin Nutr* 46:631–635.
- Scheie AA, Petersen FC. 2004. The biofilm concept: consequences for future prophylaxis of oral diseases? *Crit Rev Oral Biol Med* 15:4–12.
- Schenkein HA, Loos BG. 2013. Inflammatory mechanisms linking periodontal diseases to cardiovascular diseases. *J Clin Periodontol* 40:S51–S69.
- Schulz PD. 1977. Task activity and anterior tooth grooving in prehistoric California Indians. *Am J Phys Anthropol* 46:87–92.
- Schulz PD. 1981. *Osteoarchaeology and Subsistence Change in Prehistoric Central California*. PhD Dissertation, University of California, Davis.
- Sciulli PW. 1997. Dental evolution in prehistoric Native Americans of the Ohio Valley area. I. Wear and pathology. *Int J Osteoarchaeol* 7:507–524.
- Scott EC. 1979. Dental wear scoring technique. *Am J Phys Anthropol* 51:213–218.
- Seneviratne CJ, Yip JWY, Chang JWW, Zhang CF, Samaranyake LP. 2013. Effect of culture media and nutrients on biofilm growth kinetics of laboratory and clinical strains of *Enterococcus faecalis*. *Arch Oral Biol* 58:1327–1334.
- Serrano J, Puupponen-Pimiä R, Dauer A, Aura A, Saura-Calixto F. 2009. Tannins: current knowledge of food sources,

- intake, bioavailability and biological effects. *Mol Nutr Food Res* 53:S310–S329.
- Shanks R, Shanks LW. 2006. Indian baskets of Central California, art, culture and history. Novato, CA: Costano Books.
- Shanks R, Shanks LW. 2010. California Indian baskets. Novato, CA: Costano Books.
- Shear M. 1992. Cysts of the oral regions, 3rd ed. Oxford: Wright.
- Sheiham A, Sabbah W. 2010. Using universal patterns of caries for planning and evaluating dental care. *Caries Res* 44:141–150.
- Silva TA, Garlet GP, Fukada SY, Silva JS, Cunha FQ. 2007. Chemokines in oral inflammatory diseases: apical periodontitis and periodontal disease. *J Dent Res* 86:306–319.
- Silverstone LM, Johnson N, Hardie JM, Williams RAD. 1981. Dental caries. aetiology, pathology, and prevention. London: The Macmillan Press.
- Simón-Soro A, Belda-Ferre P, Carera-Rubio R, Alcaraz LD, Mira A. 2013. A tissue-dependent hypothesis of dental caries. *Caries Res* 47:591–600.
- Siqueira JF. 2002. Endodontic infections: concepts, paradigms, and perspectives. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 94:281–293.
- Siqueira JF, Rôças IN. 2009. Diversity of endodontic microbiota revisited. *J Dent Res* 88:969–981.
- Šlaus M, Bedić Ž, Rajić P, Šikanjić, Vodanović M, Kunić AD. 2011. Dental health at the transition from the late antique to the early medieval period on Croatia's east Adriatic coast. *Int J Osteoarchaeol* 21:577–590.
- Slots J. 2010. Human viruses in periodontitis. *Periodontol* 2000 53:89–110.
- Smith BH. 1984. Patterns of molar wear in hunter-gatherers and agriculturalists. *Am J Phys Anthropol* 63:39–56.
- Socransky SS, Haffajee AD. 2005. Periodontal microbial ecology. *Periodontol* 2000 38:135–187.
- Soni PL, Sharma H, Dun D, Gharia MM. 1993. Physicochemical properties of *Quercus leucotrichophora* (oak) starch. *Starch-Stärke* 45:127–130.
- Stabholz A, Soskolne WA, Shapira L. 2010. Genetic and environmental risk factors for chronic periodontitis and aggressive periodontitis. *Periodontol* 2000 53:138–153.
- Stevenson DG, Jane J, Inglett GE. 2006. Physicochemical properties of pin oak (*Quercus palustris* Muenchh.) acorn starch. *Starch-Stärke* 58:553–560.
- Takahashi N, Nyvad B. 2011. The role of bacteria in the caries process: ecological perspectives. *J Dent Res* 90:294–303.
- Tal H. 1985. Periodontal bone loss in dry mandibles of South African blacks: a biometric study. *J Dent Res* 64:925–929.
- Taubman MA, Valverde P, Han X, Kawai T. 2005. Immune response: the key to bone resorption in periodontal disease. *J Periodontol* 76:2033–2041.
- Temple DH, Larsen CS. 2007. Dental caries as evidence for agriculture and subsistence variation during the Yayoi period in prehistoric Japan: biocultural interpretations of an economy in transition. *Am J Phys Anthropol* 134:501–512.
- Thomson WM, Shearer DM, Broadbent JM, Foster Page LA, Poulton R. 2013. The natural history of periodontal attachment loss during the third and fourth decades of life. *J Clin Periodontol* 40:672–680.
- Thurnheer T, Giertsen E, Gmür, Guggenheim B. 2008. Cariogenicity of soluble starch in oral in vitro biofilm and experimental rat caries studies: a comparison. *J Appl Microbiol* 105: 829–836.
- Tomenchuk J, Mayhall JT. 1979. A correlation of tooth wear and age among modern Igloodik Eskimos. *Am J Phys Anthropol* 51:67–78.
- Torobinejad M, Bakland LK. 1980. Prostaglandins: their possible role in the pathogenesis of pulpal and periapical diseases. *J Endocrinol* 6:733–739.
- Tushingham S, Bettinger RL. 2013. Why foragers choose acorns before salmon: storage, mobility, and risk in aboriginal California. *J Anthropol Archaeol* 32:527–537.
- Varrela TM, Paunio K, Wouters FR, Tiekso J, Söder P-Ö. 1995. The relation between tooth eruption and alveolar crest height in a human skeletal sample. *Arch Oral Biol* 40:175–180.
- Vlot AC, Dempsey DA, Klessig DF. 2009. Salicylic acid, a multifaceted hormone to combat disease. *Annu Rev Phytopathol* 47:177–206.
- Vodanović M, Peroš K, Zukanović A, Knežević M, Novak M, Šlaus M, Brkić H. 2012. Periodontal disease at the transition from the late antique to the early mediaeval period in Croatia. *Arch Oral Biol* 57:1362–1376.
- Walker PL. 1978. A quantitative analysis of dental attrition rates in the Santa Barbara Channel area. *Am J Phys Anthropol* 48:101–106.
- Walker PL. 2006. Skeletal biology: California. In: Ubelaker DH, editor. *Handbook of North American Indians, Vol. 3: Environment, origins, and population*. Washington, D.C.: Smithsonian Institution. p 548–556.
- Walker PL, Erlandson JM. 1986. Dental evidence for prehistoric dietary change on the northern Channel Islands, California. *Am Antiqu* 51:375–383.
- Wasterlain SN, Cunha E, Hillson S. 2011. Periodontal disease in a Portuguese identified skeletal sample from the late nineteenth and early twentieth centuries. *Am J Phys Anthropol* 145:30–42.
- Wasterlain SN, Hillson S, Cunha E. 2009. Dental caries in a Portuguese identified skeletal sample from the late 19th and early 20th centuries. *Am J Phys Anthropol* 140:64–79.
- Watson JT. 2008. Prehistoric dental disease and the dietary shift from cactus to cultigens in northwest Mexico. *Int J Osteoarchaeol* 18:202–212.
- West GJ, Davis OK, Wallace WJ. 1991. Fluted points at Tulare Lake, California: environmental background. In: Wallace WJ, Riddell FA, editors. *Background to a study of Tulare Lake's archaeological past*. Salinas, CA: Coyote Press. p 1–10.
- Whittaker DK, Molleson T, Daniel AT, Williams JT, Rose P, Resteghini R. 1985. Quantitative assessment of tooth wear, alveolar crest height and continuing eruption in a Romano-British population. *Arch Oral Biol* 30:493–501.
- Wiberg R, editor. 2010. *Archaeological investigations at CA-CCO-18/548: final report for the Vineyards at Marsh Creek Project, Contra Costa County, California*. San Francisco: Holman and Associates Archaeological Consultants.
- Williams JS, Murphy MS. 2013. Living and dying as subjects of the Inca Empire: adult diet and health at Puruchuco-Huaquerones, Peru. *J Anthropol Archaeol* 32:165–179.
- Wilson M. 2005. *Microbial inhabitants of humans. Their ecology and role in health and disease*. Cambridge, UK: Cambridge University Press.
- Wohlgenuth E. 1996. Resource intensification in prehistoric central California: evidence from archaeobotanical data. *J Calif Great Basin Anthropol* 18:81–103.
- Wohlgenuth E. 2010. Plant remains. In: Wiberg R, editor. *Archaeological investigations at CA-CCO-18/548: final report for the Vineyards at Marsh Creek Project, Contra Costa County, California*. San Francisco: Holman and Associates Archaeological Consultants. p 275–285.
- Yoo S, Lee C, Kim B, Shin M. 2012. The properties and molecular structures of gusiljatbam starch compared to those of acorn and chestnut starches. *Starch-Stärke* 64:339–347.